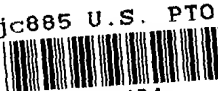


JUL 18 2001

09/889599

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE
REQUEST FOR FILING NATIONAL PHASE OF
PCT APPLICATION UNDER 35 U.S.C. 371 AND 37 CFR 1.494 OR 1.495

To: Hon. Commissioner of Patents
Washington, D.C. 20231



07/18/01



00909

TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)

Atty Dkt: P 0281500 /Z 70472/UST
M# /Client Ref.

From: Pillsbury Winthrop LLP, IP Group:

Date: July 18, 2001

This is a **REQUEST** for **FILING** a PCT/USA National Phase Application based on:

1. International Application	2. International Filing Date	3. Earliest Priority Date Claimed
<u>PCT/GB00/00265</u> ↑ country code	<u>31</u> <u>January</u> <u>2000</u> Day MONTH Year	<u>5</u> <u>February</u> <u>1999</u> Day MONTH Year (use item 2 if no earlier priority)

Measured from the earliest priority date in item 3, this PCT/USA National Phase Application Request is being filed within:

(a) ☐ 20 months from above item 3 date (b) ☒ 30 months from above item 3 date,

(c) Therefore, the due date (unextendable) is August 5, 2001

Title of Invention INDOLE DERIVATIVES AND THEIR USE AS MCP-1 ANTAGONISTS

Inventor(s) FAULL, Alan Wellington et al

Applicant herewith submits the following under 35 U.S.C. 371 to effect filing:

7. ☒ Please immediately start national examination procedures (35 U.S.C. 371 (f)).
8. ☒ **A copy of the International Application** as filed (35 U.S.C. 371(c)(2)) is transmitted herewith (file if in English but, if in foreign language, file only if not transmitted to PTO by the International Bureau) including:
- a. ☒ Request;
 - b. ☒ Abstract;
 - c. 46 pgs. Spec. and Claims;
 - d. sheet(s) Drawing which are ☐ informal ☐ formal of size ☒ A4 ☐ 11"
9. ☒ **A copy of the International Application has been transmitted by the International Bureau.**
10. **A translation of the International Application** into English (35 U.S.C. 371(c)(2))
- a. ☐ is transmitted herewith including: (1) ☐ Request; (2) ☐ Abstract;
(3) pgs. Spec. and Claims;
(4) sheet(s) Drawing which are:
☐ informal ☐ formal of size ☐ A4 ☐ 11"
 - b. ☐ is not required, as the application was filed in English.
 - c. ☐ is not herewith, but will be filed when required by the forthcoming PTO Missing Requirements Notice per Rule 494(c) if box 4(a) is X'd or Rule 495(c) if box 4(b) is X'd.
 - d. ☐ Translation verification attached (not required now).

09/889599

JC18 Rec'd PCT/PTO 1 8 JUL 2001

11. ☒ Please see the attached Preliminary Amendment
12. ☐ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)), i.e., **before 18th month from first priority date above in item 3, are transmitted herewith (file only if in English) including:**
13. ☒ PCT Article 19 claim amendments (if any) have been transmitted by the International Bureau
14. ☐ Translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)), i.e., of **claim amendments made before 18th month, is attached (required by 20th month from the date in item 3 if box 4(a) above is X'd, or 30th month if box 4(b) is X'd, or else amendments will be considered canceled).**
15. **A declaration of the inventor** (35 U.S.C. 371(c)(4))
- a. ☐ is submitted herewith ☐ Original ☐ Facsimile/Copy
- b. ☒ is not herewith, but will be filed when required by the forthcoming PTO Missing Requirements Notice per Rule 494(c) if box 4(a) is X'd or Rule 495(c) if box 4(b) is X'd.

16. **An International Search Report (ISR):**

- a. Was prepared by ☒ European Patent Office ☐ Japanese Patent Office ☐ Other
- b. ☒ has been transmitted by the international Bureau to PTO.
- c. ☒ copy herewith (2 pg(s).) ☒ plus Annex of family members (1 pg(s).).

International Preliminary Examination Report (IPER):

- a. ☒ has been transmitted (if this letter is filed after 28 months from date in item 3) in English by the International Bureau with Annexes (if any) in original language.
- b. ☒ copy herewith in English.
- c.1 ☐ IPER Annex(es) in original language ("Annexes" are amendments made to claims/spec/drawings during Examination) including attached amended:
- c.2 ☐ Specification/claim pages # _____ claims # _____
Dwg Sheets # _____
- d. ☐ Translation of Annex(es) to IPER **(required by 30th month due date, or else annexed amendments will be considered canceled).**

18. **Information Disclosure Statement** including:

- a. ☒ Attached Form PTO-1449 listing documents
- b. ☐ Attached copies of documents listed on Form PTO-1449
- c. ☒ A concise explanation of relevance of ISR references is given in the ISR.

19. ☐ **Assignment** document and Cover Sheet for recording are attached. Please mail the recorded assignment document back to the person whose signature, name and address appear at the end of this letter.

20. ☐ Copy of Power to IA agent.

21. ☐ **Drawings** (complete only if 8d or 10a(4) not completed): _____ sheet(s) per set: ☐ 1 set informal;
☐ Formal of size ☐ A4 ☐ 11"

22. Small Entity Status ☒ is **Not** claimed ☐ is claimed (**pre-filing confirmation required**)
- 22(a) _____ (No.) Small Entity Statement(s) enclosed (since 9/8/00 Small Entity Statements(s) not essential to make claim)

23. **Priority** is hereby claimed under 35 U.S.C. 119/365 based on the priority claim and the certified copy, both filed in the International Application during the international stage based on the filing in (country) **GREAT BRITAIN** of:

	Application No.	Filing Date		Application No.	Filing Date
(1)	9902461.4	February 5, 1999	(2)	_____	_____
(3)	_____	_____	(4)	_____	_____
(5)	_____	_____	(6)	_____	_____

- a. ☒ See Form PCT/IB/304 sent to US/DO with copy of priority documents. If copy has not been received, please proceed promptly to obtain same from the IB.
- b. ☐ Copy of Form PCT/IB/304 attached.

24. Attached: 2 copies of Form PCT/IB/306

09/889599
JC13 Rec'd PCT/PTO 1 8 JUL 200125 Per Item 17.c2, **cancel original** pages # _____, claims # _____, Drawing Sheets # _____**26. Calculation of the U.S. National Fee (35 U.S.C. 371 (c)(1)) and other fees is as follows:**Based on amended claim(s) per above item(s) ☐ 12, ☐ 14, ☐ 17, ☐ 25 (hilit) ☐

Total Effective Claims	minus 20 =	x \$18/\$9 =	\$0	966/967
Independent Claims	minus 3 =	x \$80/\$40 =	\$0	964/965
If any proper (ignore improper) Multiple Dependent claim is present,		add \$270/\$135	+0	968/969

BASIC NATIONAL FEE (37 CFR 1.492(a)(1)-(4)): **→→ BASIC FEE REQUIRED, NOW →→→→**A. If country code letters in item 1 are **not** "US", "BR", "BB", "TT", "MX", "IL", "NZ", "IN" or "ZA"

See item 16 re:

1. Search Report was <u>not</u> prepared by EPO or JPO -----	add \$1000/\$500	960/961
2. Search Report was prepared by EPO or JPO -----	add \$860/\$430 +860	970/971

SKIP B, C, D AND E UNLESS country code letters in item 1 are "US", "BR", "BB", "TT", "MX", "IL", "NZ", "IN" or "ZA"

→ <input type="checkbox"/> B. If USPTO did not issue both International Search Report (ISR) and (if box 4(b) above is X'd) the International Examination Report (IPER), -----	add \$1000/\$500 +0	960/961
→ <input type="checkbox"/> C. If USPTO issued ISR but not IPER (or box 4(a) above is X'd), -----	add \$710/\$355 +0	958/959
→ <input type="checkbox"/> D. If USPTO issued IPER but IPER Sec. V boxes <u>not</u> all 3 YES, -----	add \$690/\$345 +0	956/957
→ <input type="checkbox"/> E. If international preliminary examination fee was paid to USPTO and Rules 492(a)(4) and 496(b) <u>satisfied</u> (IPER Sec. V <u>all</u> 3 boxes YES for <u>all</u> claims), -----	add \$100/\$50 +0	962/963

27. SUBTOTAL =	\$860	
28. If Assignment box 19 above is X'd, add Assignment Recording fee of ----\$40	+0	(581)
29. Attached is a check to cover the -----	TOTAL FEES	\$860

Our Deposit Account No. 03-3975

Our Order No. 009901 | 0281500
C# M#

00909

CHARGE STATEMENT: The Commissioner is hereby authorized to charge any fee specifically authorized hereafter, or any missing or insufficient fee(s) filed, or asserted to be filed, or which should have been filed herewith or concerning any paper filed hereafter, and which may be required under Rules 16-18 and 492 (missing or insufficient fee only) now or hereafter relative to this application and the resulting Official document under Rule 20, or credit any overpayment, to our Account/Order Nos. shown above for which purpose a duplicate copy of this sheet is attached.This CHARGE STATEMENT does not authorize charge of the issue fee until/unless an issue fee transmittal form is filed**Pillsbury Winthrop LLP**
Intellectual Property GroupBy Atty: Donald J. BirdReg. No. 25323

Sig:

Fax: (703) 905-2500
Tel: (703) 905-2018

Atty/Sec: DJB/mhn

NOTE: File in duplicate with 2 postcard receipts (PAT-103) & attachments.

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re PATENT APPLICATION OF

Inventor(s): FAULL, Alan Wellington et al

Filed: Herewith

Title: INDOLE DERIVATIVES AND THEIR USE AS MCP-1 ANTAGONISTS

July 18, 2001

PRELIMINARY AMENDMENT

Hon. Commissioner of Patents
Washington, D.C. 20231

Sir:

Please amend this application as follows:

IN THE SPECIFICATION:

At the top of the first page, just under the title, insert

☒ --This application is the National Phase of International Application
PCT/GB00/00265 filed January 31, 2000 which designated the U.S.

and that International Application

☒ was ☐ was not published under PCT Article 21(2) in English.--

Respectfully submitted,

PILLSBURY WINTHROP LLP
Intellectual Property Group

By: 

Attorney: Donald J. Bird

Reg. No: 25323

Tel. No.: (703) 905-2018

Fax No.: (703) 905-2500

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1600 Tysons Boulevard

McLean, VA 22102
(703) 905-2000

INDOLE DERIVATIVES AND THEIR USE AS MCP-1 ANTAGONISTS

The present invention relates to anti-inflammatory compounds that act via antagonism of the CCR2 receptor, (also known as the MCP-1 receptor), leading *inter alia* to inhibition of Monocyte Chemoattractant Protein-1 (MCP-1). These compounds contain an indole moiety. The invention further relates to pharmaceutical compositions containing them, processes for their preparation, intermediates useful in their preparation and to their use as therapeutic agents.

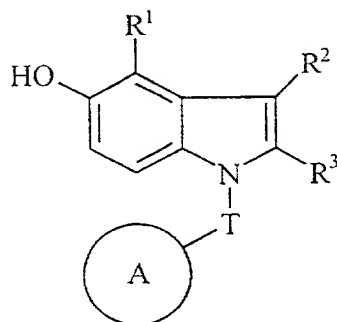
MCP-1 is a member of the chemokine family of pro-inflammatory proteins which mediate leukocyte chemotaxis and activation. MCP-1 is a C-C chemokine which is one of the most potent and selective T-cell and monocyte chemoattractant and activating agents known. MCP-1 has been implicated in the pathophysiology of a large number of inflammatory diseases including rheumatoid arthritis, glomerular nephritides, lung fibrosis, restenosis (International Patent Application WO 94/09128), alveolitis (Jones et al., 1992, *J. Immunol.*, **149**, 2147) and asthma. Other disease areas where MCP-1 is thought to play a part in their pathology are atherosclerosis (e.g. Koch et al., 1992, *J. Clin. Invest.*, **90**, 772 -779), psoriasis (Deleuran et al., 1996, *J. Dermatological Science*, **13**, 228-236), delayed-type hypersensitivity reactions of the skin, inflammatory bowel disease (Grimm et al., 1996, *J. Leukocyte Biol.*, **59**, 804-812), multiple sclerosis and brain trauma (Berman et al, 1996, *J. Immunol.*, **156**, 3017-3023). An MCP-1 inhibitor may also be useful to treat stroke, reperfusion injury, ischemia, myocardial infarction and transplant rejection.

MCP-1 acts through the CCR2 receptor. MCP-2 and MCP-3 may also act, at least in part, through this receptor. Therefore in this specification, when reference is made to "inhibition or antagonism of MCP-1" or "MCP-1 mediated effects" this includes inhibition or antagonism of MCP-2 and/or MCP-3 mediated effects when MCP-2 and/or MCP-3 are acting through the CCR2 receptor.

The applicants have found a class of compounds containing an indole moiety which have useful inhibitory activity against MCP-1. Co-pending application UK 9716657.3 discloses a class of indoles with MCP-1 inhibitory activity. This application is based on the surprising discovery that particular substituted 5-hydroxy

indoles are MCP-1 inhibitors which possess unexpected and beneficial properties with respect to potency and/or blood levels and/or bioavailability and/or solubility.

Accordingly, the present invention provides a compound of the formula (I):



(I)

wherein:

R¹ is hydrogen, halo or methoxy;

R² is hydrogen, halo, methyl, ethyl or methoxy;

R³ is carboxy, tetrazolyl or -CONHSO₂R⁴ where R⁴ is methyl, ethyl, phenyl,

2,5-dimethylisoxazolyl or trifluoromethyl;

T is -CH₂- or -SO₂-; and

ring A is 3-chlorophenyl, 4-chlorophenyl, 3-trifluoromethylphenyl,

3,4-dichlorophenyl, 3,4-difluorophenyl, 3-fluoro-4-chlorophenyl,

3-chloro-4-fluorophenyl or 2,3-dichloropyrid-5-yl;

or a pharmaceutically acceptable salt or prodrug thereof.

In this specification the term "alkyl" includes both straight and branched chain alkyl groups but references to individual alkyl groups such as "propyl" are specific for the straight chain version only. The term "halo" refers to fluoro, chloro, bromo and iodo.

Particular novel compounds of the invention include, for example, compounds of the formula (I), or pharmaceutically-acceptable salts or prodrugs thereof, wherein, unless otherwise stated:

a) R¹ has any of the values defined in i) - iii) hereinafter or a combination of two of these values;

b) R² has any of the values defined in iv) - viii) hereinafter or a combination of two of these values;

c) R^3 has any of the values defined in ix) - xi) hereinafter or a combination of two of these values;

e) T has any of the values defined in xii) - xiii) hereinafter;

f) ring A has any of the values defined in xiv) - xxi) hereinafter or a combination of

5 two or more of these values;

i) R^1 is hydrogen;

ii) R^1 is halo;

iii) R^1 is methoxy;

iv) R^2 is hydrogen;

10 v) R^2 is halo;

vi) R^2 is methyl;

vii) R^2 is ethyl;

viii) R^2 is methoxy;

ix) R^3 is carboxy;

15 x) R^3 is tetrazolyl;

xi) R^3 is $-\text{CONHSO}_2\text{R}^4$ where R^4 is methyl, ethyl, phenyl, 2,5-dimethylisoxazolyl or trifluoromethyl;

xii) T is $-\text{CH}_2-$;

xiii) T is $-\text{SO}_2-$;

20 xiv) Ring A is 3-chlorophenyl;

xv) Ring A is 4-chlorophenyl;

xvi) Ring A is 3-trifluoromethylphenyl;

xvii) Ring A is 3,4-dichlorophenyl;

xviii) Ring A is 3,4-difluorophenyl;

25 xix) Ring A is 3-fluoro-4-chlorophenyl;

xx) Ring A is 3-chloro-4-fluorophenyl; and

xxi) Ring A is 2,3-dichloropyrid-5-yl.

Preferably R^1 is hydrogen.

Preferably R^2 is hydrogen.

30 Preferably R^3 is carboxy.

Preferably T is $-\text{CH}_2-$.

Preferably Ring A is 3-chlorophenyl, 4-chlorophenyl, 3-trifluoromethylphenyl, 3,4-dichlorophenyl, 3,4-difluorophenyl, 3-fluoro-4-chlorophenyl or 3-chloro-4-fluorophenyl.

More preferably Ring A is 3,4-dichlorophenyl, 3-fluoro-4-chlorophenyl or 3-chloro-4-fluorophenyl.

For example, Ring A is 3,4-dichlorophenyl or 3-chloro-4-fluorophenyl.

In another aspect of the invention preferably Ring A is 3,4-dichlorophenyl, 2,3-dichloropyrid-5-yl or 3-chloro-4-fluorophenyl.

Therefore, in a preferred aspect of the invention there is provided a compound of formula (I) as depicted above wherein:

R¹ is hydrogen;

R² is hydrogen;

R³ is carboxy;

T is -CH₂-; and

Ring A is 3,4-dichlorophenyl, 3-fluoro-4-chlorophenyl or 3-chloro-4-fluorophenyl, in particular 3,4-dichlorophenyl or 3-chloro-4-fluorophenyl; or a pharmaceutically acceptable salt or prodrug thereof.

Preferred compounds of the invention include any one of the Examples. More preferred compounds of the invention are Examples 1, 3 and 4, for instance, Example 1 and 3.

The invention further relates to all tautomeric forms of the compounds of formula (I).

It is also to be understood that certain compounds of the formula (I) can exist in solvated as well as unsolvated forms such as, for example, hydrated forms. It is to be understood that the invention encompasses all such solvated forms.

Compounds of formula (I) are inhibitors of monocyte chemoattractant protein-1. In addition, they appear to inhibit RANTES induced chemotaxis. RANTES (Regulated upon Activation, Normal T-cell Expressed and Secreted) is another chemokine from the same family as MCP-1, with a similar biological profile, but acting through the CCR1 receptor. As a result, these compounds can be used to treat disease mediated by these agents, in particular inflammatory disease.

Suitable pharmaceutically acceptable salts of compounds of formula (I) include base salts such as an alkali metal salt for example sodium, an alkaline earth metal salt for example calcium or magnesium, an organic amine salt for example triethylamine, morpholine, *N*-methylpiperidine, *N*-ethylpiperidine, procaine, 5 dibenzylamine, *N,N*-dibenzylethylamine or amino acids for example lysine. In another aspect, where the compound is sufficiently basic, suitable salts include acid addition salts such as methanesulphonate, fumarate, hydrochloride, hydrobromide, citrate, maleate and salts formed with phosphoric and sulphuric acid. There may be more than one cation or anion depending on the number of charged functions and the valency of 10 the cations or anions. A preferred pharmaceutically acceptable salt is a sodium salt.

Various forms of prodrugs are known in the art. For examples of such prodrug derivatives, see:

- a) Design of Prodrugs, edited by H. Bundgaard, (Elsevier, 1985) and Methods in Enzymology, Vol. 42, p. 309-396, edited by K. Widder, *et al.* (Academic Press, 15 1985);
- b) A Textbook of Drug Design and Development, edited by Krogsgaard-Larsen and H. Bundgaard, Chapter 5 "Design and Application of Prodrugs", by H. Bundgaard p. 113-191 (1991);
- 20 c) H. Bundgaard, Advanced Drug Delivery Reviews, 8, 1-38 (1992);
- d) H. Bundgaard, *et al.*, Journal of Pharmaceutical Sciences, 77, 285 (1988); and
- e) N. Kakeya, *et al.*, Chem Pharm Bull, 32, 692 (1984).

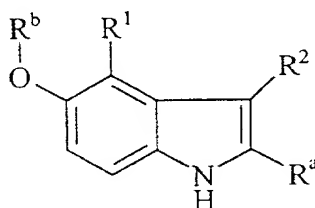
Examples of such prodrugs are *in vivo* cleavable esters of a compound of 25 the invention. An *in vivo* cleavable ester of a compound of the invention containing a carboxy group is, for example, a pharmaceutically-acceptable ester which is cleaved in the human or animal body to produce the parent acid. Suitable pharmaceutically-acceptable esters for carboxy include C₁₋₆alkyl esters, for example methyl or ethyl; C₁₋₆alkoxymethyl esters, for example methoxymethyl; 30 C₁₋₆alkanoyloxymethyl esters, for example pivaloyloxymethyl; phthalidyl esters; C₃₋₈cycloalkoxycarbonyloxyC₁₋₆alkyl esters, for example

1-cyclohexylcarbonyloxyethyl; 1,3-dioxolan-2-ylmethyl esters, for example 5-methyl-1,3-dioxolan-2-ylmethyl; C₁₋₆alkoxycarbonyloxyethyl esters, for example 1-methoxycarbonyloxyethyl; aminocarbonylmethyl esters and mono- or di-N-(C₁₋₆alkyl) versions thereof, for example N,N-dimethylaminocarbonylmethyl esters and N-ethylaminocarbonylmethyl esters; and may be formed at any carboxy group in the compounds of this invention. An *in vivo* cleavable ester of a compound of the invention containing a hydroxy group is, for example, a pharmaceutically-acceptable ester which is cleaved in the human or animal body to produce the parent hydroxy group. Suitable pharmaceutically acceptable esters for hydroxy include C₁₋₆alkanoyl esters, for example acetyl esters; and benzoyl esters wherein the phenyl group may be substituted with aminomethyl or N-substituted mono- or di-C₁₋₆alkyl aminomethyl, for example 4-aminomethylbenzoyl esters and 4-N,N-dimethylaminomethylbenzoyl esters.

Further examples of such prodrugs are *in vivo* cleavable amides of a compound of the invention. Examples of such *in vivo* cleavable amides include an N-C₁₋₆alkylamide and an N,N-di-(C₁₋₆alkyl)amide such as N-methyl, N-ethyl, N-propyl, N,N-dimethyl, N-ethyl-N-methyl or N,N-diethylamide

Another aspect of the present invention provides a process for preparing a compound of formula (I) or a pharmaceutically acceptable salt or prodrug thereof which process (wherein R¹, R², R³, T and Ring A are as defined for formula (I) unless otherwise stated) comprises of:

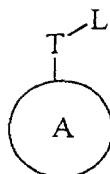
a) reacting compounds of formula (II):



(II)

where R^a is R³ or protected R³, and R^b is hydrogen or a suitable hydroxy protecting group, with a compound of formula (III):

- 7 -



(III)

where L is a displaceable group;

and thereafter if necessary:

- 5 i) converting a compound of the formula (I) into another compound of the formula (I);
- ii) removing any protecting groups; or
- iii) forming a pharmaceutically acceptable salt or prodrug thereof.

Suitable values for L are for example, a halogeno or sulphonyloxy group, for example a chloro, bromo, methanesulphonyloxy or toluene-4-sulphonyloxy group.

10 Specific reaction conditions for the above reactions are as follows.

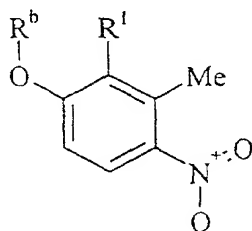
- a) Compounds of formula (II) and (III) may be reacted together in an inert solvent and a base such as *N,N*-dimethylformamide/sodium hydride or dichloromethane/sodium hydroxide or acetonitrile/potassium carbonate, or in the presence of a phase transfer catalyst such as tetra-*n*-butylammonium
- 15 hydrogensulphate. The reaction is suitably carried out for 1-6 hours preferably 1-3 hours, at a temperature of 15-30°C, preferably 20-25°C to give a compound of formula (I).

Compounds of formula (II) may be commercially available, or they may be made by modification using known processes of commercially available compounds

20 of formula (II), or they may be prepared by the following processes:

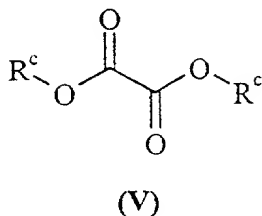
Process i)

Reacting a compound of formula (IV):



(IV)

where R^b is as defined above with a compound of formula (V)

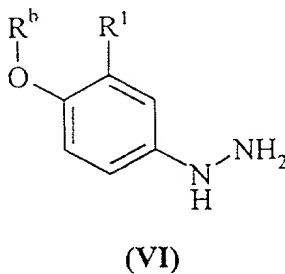


where R^c is C_{1-4} alkyl.

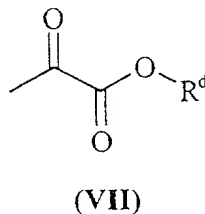
- 5 Compounds of formula (IV) and (V) are reacted together under Reissert reaction conditions such as in an inert solvent (such as tetrahydrofuran), in the presence of a base (such as potassium ethoxide), at a temperature range of 15-30°C preferably 20-25°C, for 10-20 hours preferably 15-17 hours. The resulting compound is isolated and dissolved in an alcohol such as ethanol and an organic acid (such as acetic acid) and a transition metal catalyst (such as 10% Pd/C) and cyclohexene is added. The mixture is heated at a temperature of 60-120°C preferably at 70-90°C for 15-25 hours preferably 16-20 hours to give a compound of formula (II) wherein R^a is $-CO_2C_{1-4}$ alkyl.

Process (ii)

- 15 Reacting a compound of formula (VI):



where R^b is as defined above, with a compound of formula (VII):

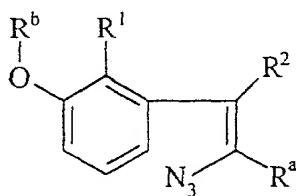


where R^d is C_{1-4} alkyl.

Compounds of formula (VI) and (VII) are reacted together under Fischer conditions such as with an organic acid (such as acetic acid), in an alcohol (such as ethanol), at a temperature of 60-90°C, preferably 75-85°C, for 1-5 hours, preferably 1-3 hours. The resulting compound is mixed with a strong acid (such as polyphosphoric acid) and heated at 90-150°C preferably 100-120°C, for 0.5-4 hours, preferably 0.5-2 hours to give a compound of formula (II) in which R² is hydrogen. Then, if desired, R² can be optionally converted into another value of R² as defined in formula (I) using techniques known in the art such as those described below.

Process (iii)

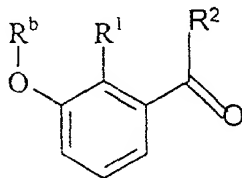
10 Cyclisation of a compound of formula (VIII)



(VIII)

where R¹, R^a, R^b and R² are as defined above.

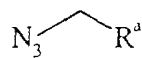
15 Cyclisation may be effected by refluxing the compound in an organic solvent such as xylene. Compounds of formula (VIII) are suitably prepared by reacting a compound of formula (IX)



(IX)

20

where R¹, R² and R^b are as defined above, with a compound of formula (X)

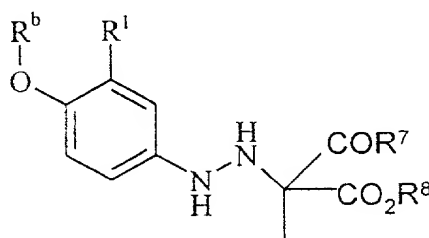


(X)

where R^a is as defined above. The reaction is suitably effected in an organic solvent such as an alcohol, in particular methanol, in the presence of a base such as an alkali metal alkoxide, in particular sodium methoxide. Moderate temperatures of from -30 to 20°C are suitably employed.

5 Process (iv)

In yet a further modification, compounds of formula (II) are prepared by cyclisation of a compound of formula (XI)

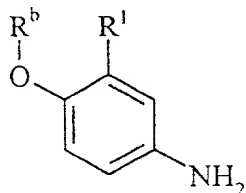


(XI)

where R^1 and R^b are as defined above, R^7 is alkyl, such as methyl, and R^8 is a carboxy protecting group such as alkyl, in particular methyl.

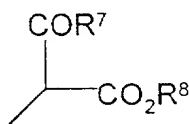
Cyclisation is suitably effected under Japp Klingemann conditions, by warming a solution of the compound in an organic solvent such as toluene and a suitable acid, such as p-toluene sulphonic acid.

Compounds of formula (XI) are suitably prepared by reacting a compound of formula (XII)



(XII)

where R^1 , R^b , R^5 and R^6 are as defined above, with a compound of formula (XIII)



(XIII)

where R^7 and R^8 are as defined in relation to formula (XI). The compound of formula (XII) is suitably dissolved in a dilute acid such as 1.5N HCl in the presence of a nitrite such as sodium nitrite at moderately low temperatures from -30 to 0°C, preferably -5°C.

- 5 This solution is then mixed with a solution of a compound of formula (XIII) in an organic solvent such as ethanol, in the presence of a solution of a base such as an alkali metal hydroxide, for example aqueous sodium hydroxide solution.

- 10 Compounds of formula (III), (IV), (V), (VI), (VII), (VIII), (X), (XII) and (XIII) are known or commercially available or are prepared by processes known in the art by standard manipulation of commercially available or known materials.

R^c and R^d are C_{1-4} alkyl. Preferably R^c and R^d are methyl or ethyl.

- 15 It will also be appreciated that in some of the reactions mentioned herein it may be necessary/desirable to protect any sensitive groups in the compounds. The instances where protection is necessary or desirable and suitable methods for protection are known to those skilled in the art. Thus, if reactants include groups such as carboxy or hydroxy it may be desirable to protect the group in some of the reactions mentioned herein.

- 20 A suitable protecting group for a hydroxy group is, for example, an acyl group, for example an alkanoyl group such as acetyl, an aroyl group, for example benzoyl, or an arylmethyl group, for example benzyl. The deprotection conditions for the above protecting groups will necessarily vary with the choice of protecting group. Thus, for example, an acyl group such as an alkanoyl or an aroyl group may be removed, for example, by hydrolysis with a suitable base such as an alkali metal hydroxide, for example lithium or sodium hydroxide. Alternatively an arylmethyl group such as a benzyl group may be removed, for example, by hydrogenation over a catalyst such as palladium-on-carbon.

- 30 A suitable protecting group for a carboxy group is, for example, an esterifying group, for example a methyl or an ethyl group which may be removed, for example, by hydrolysis with a base such as sodium hydroxide, or for example a *t*-butyl group which may be removed, for example, by treatment with an acid, for example an

organic acid such as trifluoroacetic acid, or for example a benzyl group which may be removed, for example, by hydrogenation over a catalyst such as palladium-on-carbon.

The protecting groups may be removed at any convenient stage in the synthesis using conventional techniques well known in the chemical art.

5 Some of the intermediates described herein may be novel, for example intermediates of the formula (II), and as such they are provided as a further feature of the invention.

When a pharmaceutically-acceptable salt of a compound of formula (I) is required, it may be obtained, for example, by reaction of said compound with the
10 appropriate acid (which affords a physiologically acceptable anion), or with the appropriate base (which affords a physiologically acceptable cation), or by any other conventional salt formation procedure.

According to a further aspect of the invention there is provided a pharmaceutical composition which comprises a compound of the formula (I) as defined hereinbefore or
15 a pharmaceutically acceptable salt or prodrug thereof, in association with a pharmaceutically acceptable excipient or carrier.

The compositions of the invention may be in a form suitable for oral use (for example as tablets, lozenges, hard or soft capsules, aqueous or oily suspensions, emulsions, dispersible powders or granules, syrups or elixirs), for topical use (for
20 example as creams, ointments, gels, or aqueous or oily solutions or suspensions), for administration by inhalation (for example as a finely divided powder or a liquid aerosol), for administration by insufflation (for example as a finely divided powder) or for parenteral administration (for example as a sterile aqueous or oily solution for intravenous, subcutaneous, intramuscular or intramuscular dosing or as a suppository
25 for rectal dosing).

The compositions of the invention may be obtained by conventional procedures using conventional pharmaceutical excipients, well known in the art. Thus, compositions intended for oral use may contain, for example, one or more colouring, sweetening, flavouring and/or preservative agents.

30 Suitable pharmaceutically acceptable excipients for a tablet formulation include, for example, inert diluents such as lactose, sodium carbonate, calcium

phosphate or calcium carbonate, granulating and disintegrating agents such as corn starch or algenic acid; binding agents such as starch; lubricating agents such as magnesium stearate, stearic acid or talc; preservative agents such as ethyl or propyl p-hydroxybenzoate, and anti-oxidants, such as ascorbic acid. Tablet formulations may be uncoated or coated either to modify their disintegration and the subsequent absorption of the active ingredient within the gastrointestinal track, or to improve their stability and/or appearance, in either case, using conventional coating agents and procedures well known in the art.

Compositions for oral use may be in the form of hard gelatin capsules in which the active ingredient is mixed with an inert solid diluent, for example, calcium carbonate, calcium phosphate or kaolin, or as soft gelatin capsules in which the active ingredient is mixed with water or an oil such as peanut oil, liquid paraffin, or olive oil.

Aqueous suspensions generally contain the active ingredient in finely powdered form together with one or more suspending agents, such as sodium carboxymethylcellulose, methylcellulose, hydroxypropylmethylcellulose, sodium alginate, polyvinyl-pyrrolidone, gum tragacanth and gum acacia; dispersing or wetting agents such as lecithin or condensation products of an alkylene oxide with fatty acids (for example polyoxyethylene stearate), or condensation products of ethylene oxide with long chain aliphatic alcohols, for example heptadecaethyleneoxycetanol, or condensation products of ethylene oxide with partial esters derived from fatty acids and a hexitol such as polyoxyethylene sorbitol monooleate, or condensation products of ethylene oxide with long chain aliphatic alcohols, for example heptadecaethyleneoxycetanol, or condensation products of ethylene oxide with partial esters derived from fatty acids and a hexitol such as polyoxyethylene sorbitol monooleate, or condensation products of ethylene oxide with partial esters derived from fatty acids and hexitol anhydrides, for example polyethylene sorbitan monooleate. The aqueous suspensions may also contain one or more preservatives (such as ethyl or propyl p-hydroxybenzoate, anti-oxidants (such as ascorbic acid), colouring agents, flavouring agents, and/or sweetening agents (such as sucrose, saccharine or aspartame).

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Suppository formulations may be prepared by mixing the active ingredient with a suitable non-irritating excipient which is solid at ordinary temperatures but liquid at the rectal temperature and will therefore melt in the rectum to release the drug. Suitable excipients include, for example, cocoa butter and polyethylene glycols.

5 Topical formulations, such as creams, ointments, gels and aqueous or oily solutions or suspensions, may generally be obtained by formulating an active ingredient with a conventional, topically acceptable, vehicle or diluent using conventional procedure well known in the art.

10 Compositions for administration by insufflation may be in the form of a finely divided powder containing particles of average diameter of, for example, 30 μ or much less, the powder itself comprising either active ingredient alone or diluted with one or more physiologically acceptable carriers such as lactose. The powder for insufflation is then conveniently retained in a capsule containing, for example, 1 to 50mg of active ingredient for use with a turbo-inhaler device, such as is used for insufflation of the
15 known agent sodium cromoglycate.

20 Compositions for administration by inhalation may be in the form of a conventional pressurised aerosol arranged to dispense the active ingredient either as an aerosol containing finely divided solid or liquid droplets. Conventional aerosol propellants such as volatile fluorinated hydrocarbons or hydrocarbons may be used and the aerosol device is conveniently arranged to dispense a metered quantity of active ingredient.

For further information on Formulation the reader is referred to Chapter 25.2 in Volume 5 of Comprehensive Medicinal Chemistry (Corwin Hansch; Chairman of Editorial Board), Pergamon Press 1990.

25 The amount of active ingredient that is combined with one or more excipients to produce a single dosage form will necessarily vary depending upon the host treated and the particular route of administration. For example, a formulation intended for oral administration to humans will generally contain, for example, from 0.5 mg to 2 g of active agent compounded with an appropriate and convenient amount of excipients
30 which may vary from about 5 to about 98 percent by weight of the total composition. Dosage unit forms will generally contain about 1 mg to about 500 mg of an active

ingredient. For further information on Routes of Administration and Dosage Regimes the reader is referred to Chapter 25.3 in Volume 5 of Comprehensive Medicinal Chemistry (Corwin Hansch; Chairman of Editorial Board), Pergamon Press 1990.

The size of the dose for therapeutic or prophylactic purposes of a compound of the Formula I will naturally vary according to the nature and severity of the conditions, the age and sex of the animal or patient and the route of administration, according to well known principles of medicine. As mentioned above, compounds of the Formula I are useful in treating diseases or medical conditions which are due alone or in part to the effects of farnesylation of rats.

In using a compound of the Formula I for therapeutic or prophylactic purposes it will generally be administered so that a daily dose in the range, for example, 0.5 mg to 75 mg per kg body weight is received, given if required in divided doses. In general lower doses will be administered when a parenteral route is employed. Thus, for example, for intravenous administration, a dose in the range, for example, 0.5 mg to 30 mg per kg body weight will generally be used. Similarly, for administration by inhalation, a dose in the range, for example, 0.5 mg to 25 mg per kg body weight will be used. Oral administration is however preferred.

According to a further aspect of the present invention there is provided a compound of the formula (I) or a pharmaceutically acceptable salt or prodrug thereof, as defined hereinbefore for use in a method of treatment of the human or animal body by therapy. Conveniently, the invention provides a method of treating inflammatory disease by administering a compound of formula (I) or a pharmaceutically acceptable salt or prodrug or a pharmaceutical composition thereof, as described above.

A further feature of the present invention is a compound of formula (I) and pharmaceutically acceptable salt or prodrug thereof, for use as a medicament.

Conveniently this is a compound of formula (I), or a pharmaceutically acceptable salt or prodrug thereof, for use as a medicament for antagonising an MCP-1 mediated effect in a warm-blooded animal such as a human being.

Thus according to a further aspect of the invention there is provided the use of a compound of the formula (I), or a pharmaceutically acceptable salt or prodrug thereof,

in the manufacture of a medicament for use in antagonising an MCP-1 mediated effect in a warm-blooded animal such as a human being.

According to a further feature of the invention there is provided a method of antagonising an MCP-1 mediated effect in a warm-blooded animal, such as a human being, in need of such treatment which comprises administering to said animal an effective amount of a compound of formula (I) or a pharmaceutically acceptable salt or prodrug thereof, as defined hereinbefore.

Biological Testing.

The following biological test methods, data and Examples serve to illustrate the present invention.

Abbreviations:

ATCC	American Type Culture Collection, Rockville, USA.
BCA	Bicinchroninic acid, (used, with copper sulphate, to assay protein)
BSA	Bovine Serum Albumin
DMEM	Dulbecco's modified Eagle's medium
EGTA	Ethylenebis(oxyethylenenitrilo)tetraacetic acid
FCS	Foetal calf serum
HEPES	(N-[2-Hydroxyethyl]piperazine-N'-[2-ethanesulphonic acid])
HBSS	Hank's Balanced Salt Solution
hMCP-1	Human Monocyte Chemoattractant Protein-1
PBS	Phosphate buffered saline
PCR	Polymerase chain reaction

AMPLITAQ™, available from Perkin-Elmer Cetus, is used as the source of thermostable DNA polymerase.

Binding Buffer is 50 mM HEPES, 1 mM CaCl₂, 5 mM MgCl₂, 0.5% foetal calf serum, adjusted to pH 7.2 with 1 M NaOH.

Non-Essential Amino Acids (100X concentrate) is: L-Alanine, 890 mg/l; L-Asparagine, 1320 mg/l; L-Aspartic acid, 1330 mg/l; L-Glutamic acid, 1470 mg/l; Glycine, 750 mg/l; L-Proline, 1150 mg/l and; L-Serine, 1050 mg/l.

Hypoxanthine and Thymidine Supplement (50x concentrate) is: hypoxanthine, 680 mg/l and; thymidine, 194 mg/l.

Penicillin-Streptomycin is: Penicillin G (sodium salt); 5000 units/ml;
Streptomycin sulphate, 5000 µg/ml.

Human monocytic cell line THP-1 cells are available from ATCC, accession number ATCC TIB-202.

5 Hank's Balanced Salt Solution (HBSS) was obtained from Gibco; see *Proc. Soc. Exp. Biol. Med.*, 1949, 71, 196.

Synthetic cell culture medium, RPMI 1640 was obtained from Gibco; it contains inorganic salts [$\text{Ca}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ 100 mg/l; KCl 400 mg/l; $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 100 mg/l; NaCl 6000 mg/l; NaHCO_3 2000 mg/l & Na_2HPO_4 (anhyd) 800 mg/l], D-Glucose
10 2000 mg/l, reduced glutathione 1 mg/l, amino acids and vitamins.

FURA-2/AM is 1-[2-(5-carboxyoxazol-2-yl)-6-aminobenzofuran-5-oxy]-2-(2'-amino-5'-methylphenoxy)-ethane-*N,N,N',N'*-tetraacetic acid pentaacetoxymethyl ester and was obtained from Molecular Probes, Eugene, Oregon, USA.

Blood Sedimentation Buffer contains 8.5g/l NaCl and 10g/l hydroxyethyl
15 cellulose.

Lysis Buffer is 0.15M NH_4Cl , 10mM KHCO_3 , 1mM EDTA

Whole Cell Binding Buffer is 50 mM HEPES, 1 mM CaCl_2 , 5 mM MgCl_2 , 0.5% BSA, 0.01% NaN_3 , adjusted to pH 7.2 with 1M NaOH.

Wash buffer is 50mM HEPES, 1mM CaCl_2 , 5mM MgCl_2 , 0.5% heat
20 inactivated FCS, 0.5MNaCl adjusted to pH7.2 with 1M NaOH.

General molecular biology procedures can be followed from any of the methods described in "Molecular Cloning - A Laboratory Manual" Second Edition. Sambrook, Fritsch and Maniatis (Cold Spring Harbor Laboratory, 1989).

i) Cloning and expression of hMCP-1 receptor

25 The MCP-1 receptor B (CCR2B) cDNA was cloned by PCR from THP-1 cell RNA using suitable oligonucleotide primers based on the published MCP-1 receptor sequences (Charo *et al.*, 1994, *Proc. Natl. Acad. Sci. USA*, **91**, 2752). The resulting PCR products were cloned into vector PCR-II™ (InVitrogen, San Diego, CA.). Error free CCR2B cDNA was subcloned as a Hind III-Not I fragment into the eukaryotic
30 expression vector pCDNA3 (InVitrogen) to generate pCDNA3/CC-CKR2A and pCDNA3/CCR2B respectively.

Linearised pCDNA3/CCR2B DNA was transfected into CHO-K1 cells by calcium phosphate precipitation (Wigler *et al.*, 1979, *Cell*, **16**, 777). Transfected cells were selected by the addition of Geneticin Sulphate (G418, Gibco BRL) at 1mg/ml, 24 hours after the cells had been transfected. Preparation of RNA and Northern blotting were carried out as described previously (Needham *et al.*, 1995, *Prot. Express. Purific.*, **6**, 134). CHO-K1 clone 7 (CHO-CCR2B) was identified as the highest MCP-1 receptor B expressor.

ii) Preparation of membrane fragments

CHO-CCR2B cells were grown in DMEM supplemented with 10% foetal calf serum, 2 mM glutamine, 1x Non-Essential Amino Acids, 1x Hypoxanthine and Thymidine Supplement and Penicillin-Streptomycin (at 50 µg streptomycin/ml, Gibco BRL). Membrane fragments were prepared using cell lysis/differential centrifugation methods as described previously (Siciliano *et al.*, 1990, *J. Biol. Chem.*, **265**, 19658). Protein concentration was estimated by BCA protein assay (Pierce, Rockford, Illinois) according to the manufacturer's instructions.

iii) Assay

¹²⁵I MCP-1 was prepared using Bolton and Hunter conjugation (Bolton *et al.*, 1973, *Biochem. J.*, **133**, 529; Amersham International plc]. Equilibrium binding assays were carried out using the method of Ernst *et al.*, 1994, *J. Immunol.*, **152**, 3541. Briefly, varying amounts of ¹²⁵I-labeled MCP-1 were added to 7µg of purified CHO-CCR2B cell membranes in 100 µl of Binding Buffer. After 1 hour incubation at room temperature the binding reaction mixtures were filtered and washed 5 times through a plate washer (Brandel MLR-96T Cell Harvester) using ice cold Binding Buffer. Filter mats (Brandel GF/B) were pre-soaked for 60 minutes in 0.3% polyethylenimine prior to use. Following filtration individual filters were separated into 3.5ml tubes (Sarstedt No. 55.484) and bound ¹²⁵I-labeled MCP-1 was determined (LKB 1277 Gammamaster). Cold competition studies were performed as above using 100 pM ¹²⁵I-labeled MCP-1 in the presence of varying concentrations of unlabelled MCP-1. Non-specific binding was determined by the inclusion of a 200-fold molar excess of unlabelled MCP-1 in the reaction.

Ligand binding studies with membrane fragments prepared from CHO-CCR2B cells showed that the CCR2B receptor was present at a concentration of 0.2 pmoles/mg of membrane protein and bound MCP-1 selectively and with high affinity ($IC_{50} = 110$ pM, $K_d = 120$ pM). Binding to these membranes was completely reversible and reached equilibrium after 45 minutes at room temperature, and there was a linear relationship between MCP-1 binding and CHO-CCR2B cell membrane concentration when using MCP-1 at concentrations between 100 pM and 500 pM.

Test compounds dissolved in DMSO (5 μ l) were tested in competition with 100 pM labelled MCP-1 over a concentration range (0.01-50 μ M) in duplicate using eight point dose-response curves and IC_{50} concentrations were calculated.

Compounds tested of the present invention had IC_{50} values of 50 μ M or less in the hMCP-1 receptor binding assay described herein.

b) MCP-1 mediated calcium flux in THP-1 cells

The human monocytic cell line THP-1 was grown in a synthetic cell culture medium RPMI 1640 supplemented with 10 % foetal calf serum, 6mM glutamine and Penicillin-Streptomycin (at 50 μ g streptomycin/ml, Gibco BRL). THP-1 cells were washed in HBSS (lacking Ca^{2+} and Mg^{2+}) + 1 mg/ml BSA and resuspended in the same buffer at a density of 3×10^6 cells/ml. The cells were then loaded with 1mM FURA-2/AM for 30 min at 37°C, washed twice in HBSS, and resuspended at 1×10^6 cells/ml. THP-1 cell suspension (0.9 ml) was added to a 5 ml disposable cuvette containing a magnetic stirrer bar and 2.1 ml of prewarmed (37°C) HBSS containing 1 mg/ml BSA, 1 mM $MgCl_2$ and 2 mM $CaCl_2$. The cuvette was placed in a fluorescence spectrophotometer (Perkin Elmer, Norwalk, CT) and preincubated for 4 min at 37°C with stirring. Fluorescence was recorded over 70 sec and cells were stimulated by addition of hMCP-1 to the cuvette after 10 sec. $[Ca^{2+}]_i$ was measured by excitation at 340 nm and 380 nm alternately and subsequent measurement of the intensity of the fluorescence emission at 510 nm. The ratio of the intensities of the emitted fluorescent light following excitation at 340 nm and 380 nm, (R), was calculated and displayed to give and estimate of cytoplasmic $[Ca^{2+}]$ according to the equation:-

$$[Ca^{2+}]_i = K_d \frac{(R - R_{min})}{(R_{max} - R)} (Sf2/Sb2)$$

where the K_d for FURA-2 Ca^{2+} complex at 37°C was taken to be 224nm. R_{max} is the maximal fluorescence ratio determined after addition of 10 mM Ionomycin, R_{min} is the minimal ratio determined by the subsequent addition of a Ca^{2+} free solution containing 5 mM EGTA, and Sf2/Sb2 is the ratio of fluorescence values at 380 nm excitation determined at R_{min} and R_{max} , respectively.

Stimulation of THP-1 cells with hMCP-1 induced a rapid, transient rise in $[\text{Ca}^{2+}]_i$ in a specific and dose dependent manner. Dose response curves indicated an approximate EC_{50} of 2 nM. Test compounds dissolved in DMSO (10 μl) were assayed for inhibition of calcium release by adding them to the cell suspension 10 sec prior to ligand addition and measuring the reduction in the transient rise in $[\text{Ca}^{2+}]_i$. Test compounds were also checked for lack of agonist activity by addition in place of hMCP-1.

c) hMCP-1 and RANTES mediated chemotaxis.

In vitro chemotaxis assays were performed using the human monocytic cell line THP-1. Cell migration through polycarbonate membranes was measured by enumerating those passing through either directly by Coulter counting or indirectly by use of a colourimetric viability assay measuring the cleavage of a tetrazolium salt by the mitochondrial respiratory chain (Scudiero D.A. *et al.* 1988, *Cancer Res.*, **48**, 4827-4833).

Chemoattractants were introduced into a 96-well microtitre plate which forms the lower well of a chemotaxis chamber fitted with a PVP-free 5 μm pore size polycarbonate adhesive framed filter membrane (NeuroProbe MB series, Cabin John, MD 20818, USA) according to the manufacturer's instructions. The chemoattractant was diluted as appropriate in synthetic cell culture medium, RPMI 1640 (Gibco) or supplemented with 2 mM glutamine and 0.5% BSA, or alternatively with HBSS with Ca^{2+} and Mg^{2+} without Phenol Red (Gibco) plus 0.1% BSA. Each dilution was degassed under vacuum for 30 min and was placed (400 μl) in the lower wells of the chamber and THP-1 cells (5×10^5 in 100 μl RPMI 1640 + 0.5% BSA) were incubated in each well of the upper chamber. For the inhibition of chemotaxis the chemoattractant was kept at a constant submaximal concentration determined previously (1 nM MCP-1) and added to the lower well together with the test

compounds dissolved in DMSO (final DMSO concentration < 0.05% v/v) at varying concentrations. The chamber was incubated for 2 h at 37°C under 5 % CO₂. The medium was removed from the upper wells which were then washed out with 200 µl physiological saline before opening the chamber, wiping dry the membrane surface and centrifuging the 96-well plate at 600 g for 5 min to harvest the cells. Supernatant (150 µl) was aspirated and 10 µl of cell proliferation reagent, WST-1, {4-[3-(4-iodophenyl)-2-(4-nitrophenyl)-2H-5-tetrazolio]-1,3-phenyl disulfonate} plus an electron coupling reagent (Boehringer Mannheim, Cat.no. 1644 807) was added back to the wells. The plate was incubated at 37°C for 3 h and the absorbance of the soluble formazan product was read on a microtitre plate reader at 450 nm. The data was input into a spreadsheet, corrected for any random migration in the absence of chemoattractant and the average absorbance values, standard error of the mean, and significance tests were calculated. hMCP-1 induced concentration dependent cell migration with a characteristic biphasic response, maximal 0.5-1.0 nm.

In an alternative form of the above assay, fluorescently tagged cells can be used in order to assist in end point detection. In this case, the THP-1 cells used are fluorescently tagged by incubation in the presence of 5mM Calcein AM (Glycine, N,N'-[[3',6'-bis(acetyloxy)-3-oxospiro[isobenzofuran-1(3H),9'-[9H]xanthene]-2',7'-diyl]bis(methylene)] bis[N-[2-[(acetyloxy)methoxy]-2-oxoethyl]]-bis[(acetyloxy)methyl] ester; Molecular Probes) for 45 minutes in the dark. Cells are harvested by centrifugation and resuspended in HBSS (without Phenol Red) with Ca²⁺, Mg²⁺ and 0.1% BSA. 50µl (2x10⁵ cells) of the cell suspension are placed on the filter above each well and, as above, the unit is incubated at 37°C for 2 hours under 5% CO₂. At the end of the incubation, cells are washed off the upper face of the filter with phosphate buffered saline, the filter removed from the plate and the number of cells attracted to either the underside of the filter or the lower well estimated by reading fluorescence at 485nm excitation, 538nm emission wavelengths (fmax, Molecular Devices). The data was input into a spreadsheet, corrected for any random migration in the absence of chemoattractant and the average fluorescence values, standard error of the mean, percentage inhibition and IC₅₀ of compounds under test and significance tests can be

calculated. In addition to MCP-1 induced chemotaxis, this alternative form of the assay was also used to measure inhibition of RANTES (2nM) induced chemotaxis.

d) Binding to human peripheral blood mononuclear cells(PBMCs)

i) Preparation of human PBMCs

5 Fresh human blood (200ml) was obtained from volunteer donors, collected into sodium citrate anticoagulant to give a final concentration of 0.38%. The blood was mixed with Sedimentation Buffer and incubated at 37°C for 20 minutes. The supernatant was collected and centrifuged at 1700rpm for 5 minutes (Sorvall RT6000D). The pellet obtained was resuspended in 20 ml RPMI/BSA (1mg/ml) and 4
10 x 5mls of cells were carefully layered over 4 x 5mls of Lymphoprepä (Nycomed) in 15ml centrifuge tubes. Tubes were spun at 1700rpm for 30 minutes (Sorvall RT6000D) and the resultant layer of cells was removed and transferred to 50ml Falcon tubes. The cells were washed twice in Lysis Buffer to remove any remaining red blood cells followed by 2 washes in RPMI/BSA. Cells were resuspended in 5mls
15 of Binding Buffer. Cell number was measured on a Coulter counter and additional binding buffer was added to give a final concentration of 1.25×10^7 PBMCs /ml.

ii) Assay

[125 I]MCP-1 was prepared using Bolton and Hunter conjugation (Bolton *et al.*, 1973, *Biochem. J.*, 133, 529; Amersham International plc]. Equilibrium binding
20 assays were carried out using the method of Ernst *et al.*, 1994, *J. Immunol.*, 152, 3541. Briefly, 50µl of 125 I-labeled MCP-1 (final concentration 100pM) was added to 40µl (5×10^5 cells) of cell suspension in a 96 well plate. Compounds, diluted in Whole Cell Binding Buffer from a stock solution of 10mM in DMSO were added in a final
25 volume of 5µl to maintain a constant DMSO concentration in the assay of 5%. Total binding was determined in the absence of compound. Non-specific binding was defined by the addition of 5µl cold MCP-1 to give a final assay concentration of 100nM. Assay wells were made up to a final volume of 100µl with Whole Cell Binding Buffer and the plates sealed. Following incubation at 37°C for 60 minutes the binding reaction mixtures were filtered and washed for 10 seconds using ice cold
30 Wash Buffer using a plate washer (Brandel MLR-96T Cell Harvester). Filter mats (Brandel GF/B) were pre-soaked for 60 minutes in 0.3% polyethylenimine plus 0.2%

BSA prior to use. Following filtration individual filters were separated into 3.5ml tubes (Sarstedt No. 55.484) and bound ^{125}I -labeled MCP-1 was determined (LKB 1277 Gammamaster).

Test compound potency was determined by assay in duplicate using six point dose-response curves and IC_{50} concentrations were determined.

No physiologically unacceptable toxicity was observed at the effective dose for compounds tested of the present invention.

The invention is further illustrated, but not limited by the following Examples in which the following general procedures were used unless stated otherwise.

- i) N,N-Dimethylformamide (DMF) was dried over 4Å molecular sieves. Anhydrous tetrahydrofuran (THF) was obtained from Aldrich SURESEAL™ bottles. Other commercially available reagents and solvents were used without further purification unless otherwise stated. Organic solvent extracts were dried over anhydrous MgSO_4 .
- ii) ^1H , ^{13}C and ^{19}F NMR were recorded on Bruker WM200, WM250, WM300 or WM400 instruments using $\text{DMSO}-d_6$ with Me_4Si or CCl_3F as internal standard as appropriate, unless otherwise stated. Chemical shifts are quoted in δ (ppm) and peak multiplicities are designated as follows: s, singlet; d, doublet; dd, doublet of doublets; t, triplet; dt, doublet of triplets; q, quartet; m, multiplet; br, broad.
- iii) Mass spectra were recorded on VG 12-12 quadrupole, VG 70-250 SE, VG ZAB 2-SE or a VG modified AEI/Kratos MS9 spectrometers.
- iv) For TLC analysis, Merck precoated TLC plates (silica gel 60 F254, $d = 0.25$ mm) were used.
- v) Flash chromatography was performed on silica (Merck Kieselgel: Art.9385).

Example 1

N-(3,4-Dichlorobenzyl)-5-hydroxyindole-2-carboxylic acid

Sodium hydroxide (2M, 3 ml) was added to a stirred solution of ethyl N-(3,4-dichlorobenzyl)-5-hydroxyindole-2-carboxylate (0.1 g) in THF (3 ml) and methanol (1.5 ml). The reaction was stirred at ambient temperature for 4 hours. The reaction was concentrated *in vacuo* and the residue was dissolved in water (5 ml). The solution was acidified by the addition of aqueous hydrochloric acid (2M, 4 ml)

precipitating the product as a white solid. The product was filtered, washed with water and dried in vacuo to yield the title compound (82 mg, 89%). NMR: δ 5.77 (s, 2H), 6.81 (dd, 1H), 6.89 (dd, 1H), 6.95 (d, 1H), 7.13 (s, 1H), 7.26 (d, 1H), 7.34 (d, 1H), 7.52 (d, 1H), 9.01 (s, 1H), 12.85 (s, 1H); m/z 334 (M-H⁺).

5

The procedure described in the above example were repeated using the appropriate starting ethyl indole-2-carboxylates. Thus were obtained the compounds described below.

10 **Example 2**

N-[(2,3-Dichloropyrid-5-yl)methyl]-5-hydroxyindole-2-carboxylic acid

36% yield. NMR(CD₃SOCD₃) δ 5.80 (s, 2H), 6.84 (dd, 1H), 6.96 (d, 1H), 7.14 (s, 1H), 7.23 (d, 1H), 7.73 (d, 1H), 8.06 (d, 1H); m/z 339 (M-H⁺) 337, 335.

Example 3

15 **N-(3-Chloro-4-fluorobenzyl)-5-hydroxyindole-2-carboxylic acid**

68% yield. NMR(CD₃SOCD₃) δ 5.75 (s, 2H), 6.82 (dd, 1H), 6.95 (m, 2H), 7.12 (s, 1H), 7.2 - 7.4 (m, 3H); m/z 320 (M-H⁺), 318.

Example 4

N-(4-Chloro-3-fluorobenzyl)-5-hydroxyindole-2-carboxylic acid

20 94% yield. NMR(CD₃SOCD₃) δ 5.78 (s, 2H), 6.78 (dd, 1H), 6.80 (dd, 1H), 6.96 (d, 1H), 7.03 (dd, 1H), 7.12 (s, 1H), 7.31 (d, 1H), 7.43 (t, 1H); m/z 318 (M-H⁺).

Example 5

N-(3-Chlorobenzyl)-5-hydroxyindole-2-carboxylic acid

75% yield. m/z 300 (M-H⁺).

25 **Example 6**

N-(3-Trifluoromethylbenzyl)-5-hydroxyindole-2-carboxylic acid

81% yield. m/z 334 (M-H⁺).

Example 7

N-(4-Chlorobenzyl)-5-hydroxyindole-2-carboxylic acid

30 82% yield. m/z 300 (M-H⁺).

Example 83-Bromo-N-(3, 4-Dichlorobenzyl)-5-hydroxyindole-2-carboxylic acid

95% yield. m/z 414 (M-H⁺).

Example 95 4-Bromo-N-(3, 4-dichlorobenzyl)-5-hydroxyindole-2-carboxylic acid

96% yield. NMR(CD₃SOCD₃) δ 5.78 (s, 2H), 6.86 (dd, 1H), 7.01 (d, 1H), 7.04 (s, 1H), 7.33 (s, 1H), 7.40 (d, 1H), 7.52 (d, 1H), 9.78 (s, 1H), 13.10 (bs, 1H); m/z 414 (M-H⁺).

Example 1010 N-(3, 4-Dichlorobenzyl)-5-hydroxy-3-methylindole-2-carboxylic acid

73% yield. NMR(CD₃SOCD₃) δ 2.44 (s, 3H), 5.69 (s, 2H), 6.83 (m, 2H), 6.92 (d, 1H), 7.25 (d, 1H), 7.30 (d, 1H), 7.50 (d, 1H), 9.00 (s, 1H), 12.90 (bs, 1H); m/z 350 (M-H⁺).

Example 11N-(3,4-dichlorobenzyl)-4-fluoro-5-hydroxyindole-2-carboxylic acid

15 68% yield. NMR(CD₃SOCD₃) δ 5.80(s, 2H), 6.88(m, 1H), 7.00(t, 1H), 7.20(m, 2H), 7.32(m,1H), 7.50(m, 1H), 9.25(s, 1H), 13.10(s, 1H); M/z(M-H⁺) 351.9

Example 12N-(3,4-dichlorobenzyl) 3-methoxy-5-hydroxyindole-2-carboxylic acid

20 73% yield. NMR(CD₃SOCD₃) δ 4.3 (s, 3H), 5.7 (s, 2H), 6.9 (m, 2H), 7.1-7.4 (m, 4H); m/z 364,366(M-H⁺)

Example 13N-(3,4-dichlorobenzyl)-3-chloro-5-hydroxyindole-2-carboxylic acid

25 97% yield. NMR(CD₃SOCD₃) δ 5.75 (s, 2H), 6.9 (m, 3H), 7.3 (s, 1H), 7.45 (d, 1H), 7.5 (d, 1H), 9.35 (s, 1H); m/z 368 (M-H⁺).

Example 14N-(3,4-dichlorobenzyl)-4-chloro-5-hydroxyindole-2-carboxylic acid

30 83% yield. NMR(CD₃SOCD₃) δ 5.79 (s, 2H), 6.88 (dd, 1H), 7.01 (d, 1H), 7.11 (s, 1H), 7.3 (d,1H), 7.38 (d, 1H), 7.51 (d, 1H), 9.67 (bs, 1H); m/z 368.2 (M-H⁺)

Preparation of Starting Materials

The starting materials for the Examples above are either commercially available or are readily prepared by standard methods from known materials. For example the following reactions (Methods A-J) are illustrations but not limitations of the preparation of the starting materials used in the above reactions.

Method A

3-Chloro-4-fluorobenzyl bromide

A solution of 3-chloro-4-fluorobenzaldehyde (3 g) in THF (40 ml) was added over 2 minutes to a stirred suspension of sodium borohydride (1.07 g) in methanol (40 ml) at 0°C. The mixture was allowed to warm to room temperature and then quenched with water. The resulting suspension was partitioned between water and diethyl ether and the combined organic extracts were dried and concentrated *in vacuo*. The residue was dissolved in dichloromethane (90 ml) and triphenylphosphine (4.62 g) and tetrabromomethane (6.64 g) were added at 0°C. The mixture was allowed to warm to room temperature overnight then concentrated *in vacuo* and the residue purified by column chromatography using *iso*-hexane as eluent to yield the desired product (3.57 g, 85%). NMR: δ 4.7 (s, 2H), 7.4 (m, 2H), 7.7 (m, 1H).

In a similar manner but starting from 3-fluoro-4-chlorobenzaldehyde was prepared :-

3-Fluoro-4-chlorobenzyl bromide

74% yield. NMR: δ 4.5(s, 2H), 7.1(t, 1H), 7.25(m, 1H), 7.45(dd, 1H).

Method B

2,3-Dichloro-5-(hydroxymethyl)pyridine

Borane-tetrahydrofuran complex (1M solution in tetrahydrofuran, 52 ml) was added to a stirred solution of 5,6-dichloronicotinic acid (2 g) in tetrahydrofuran (60 ml) over 20 minutes at 0°C. The reaction mixture was allowed to warm to room temperature over 90 minutes and then cooled to 0°C and quenched with water (100 ml). The solution was saturated with solid sodium chloride and extracted with ethyl acetate and the combined organic extracts were dried and concentrated *in vacuo*. The residue was triturated with dichloromethane-50% ethyl acetate and the solid

by-product was removed by filtration. The filtrate was concentrated *in vacuo* and purified by column chromatography using isohexane / ethyl acetate (1:1 v/v) as eluent to yield the product as a white solid (820 mg, 45%). NMR: δ 4.55 (d, 2H), 5.5 (t, 1H), 8.0 (m, 1H), 8.3 (m, 1H); m/z 178.1 ($M+H^+$).

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Method C

2,3-Dichloro-5-(bromomethyl)pyridine

2,3-Dichloro-5-(hydroxymethyl)pyridine (275 mg) was dissolved in dichloromethane (10 ml) and stirred in the presence of triphenylphosphine (444 mg) and tetrabromomethane (641 mg) overnight. The solution was concentrated *in vacuo* and the residue purified by column chromatography using isohexane : 2.5% ethyl acetate as eluent to yield the product as a white solid (270 mg, 73%). NMR: δ 4.75 (s, 2H), 8.25 (m, 1H), 8.5 (m, 1H); m/z 242 ($M+H^+$).

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Method D

Ethyl 5-acetoxy-N-(3,4-dichlorobenzyl)indole-2-carboxylate

i) Ethyl 5-hydroxyindole-2-carboxylate

Boron tribromide (64.58 g) was added dropwise to a stirred solution of ethyl 5-methoxyindole-2-carboxylate (20 g) in dichloromethane (1000 ml) at -78°C under an atmosphere of argon. The reaction was allowed to warm to room temperature and stirred for a further 2 hours. The reaction was poured into ice / saturated aqueous sodium hydrogen carbonate solution with stirring and extracted with ethyl acetate. Combined organic extracts were washed with saturated aqueous sodium hydrogen carbonate solution, water, aqueous saturated sodium chloride solution and dried. The solution was concentrated *in vacuo* and the residue was purified by column chromatography using 0 - 60% diethyl ether: *iso*-hexane as eluent to yield product as a white solid (9.02 g, 48%). NMR: δ 1.31 (t, 3H), 4.29 (q, 2H), 6.79 (dd, 1H), 6.90 (dd, 1H), 7.22 (d, 1H), 8.84 (s, 1H), 11.52 (brs, 1H); m/z 206 ($M+H^+$).

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ii) Ethyl 5-acetoxyindole-2-carboxylate

A stirred solution of ethyl 5-hydroxyindole-2-carboxylate (7.79 g) and 4-dimethylaminopyridine (20 mg) in acetic anhydride (80 ml) was heated at 80°C for

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4 hours. The reaction was concentrated *in vacuo* and the residue was dissolved in ethyl acetate. Combined organic extracts were washed with hydrochloric acid (2 M), saturated aqueous sodium hydrogen carbonate solution, water, aqueous saturated sodium chloride solution and dried. The solution was concentrated *in vacuo* to yield the product as a yellow solid (9.39 g, 100 %). NMR: δ 1.20 (t, 3H), 2.10 (s, 3H), 4.19 (q, 2H), 6.86 (dd, 1H), 6.97 (d, 1H), 7.20 (s, 1H), 7.29 (d, 1H); m/z 248 (M+H⁺).

iii) Ethyl 5-acetoxy-N-(3,4-dichlorobenzyl)indole-2-carboxylate

3,4-Dichlorobenzyl bromide (5.96 g) was added to a stirred solution of ethyl 5-acetoxyindole-2-carboxylate (5.4 g) and potassium carbonate (6.94 g) in acetonitrile (500 ml) under an atmosphere of argon. The reaction was heated at 80°C for 16 hours, then concentrated *in vacuo* and the residue partitioned between ethyl acetate and water. Combined organic extracts were washed with water, saturated aqueous sodium chloride and dried. The solvent was removed *in vacuo* and the residue was triturated with *iso*-hexane to yield the product as a cream solid (5.55 g, 63%). NMR: δ 1.27 (t, 3H), 2.27 (s, 3H), 4.28 (q, 2H), 5.82 (s, 2H), 6.90 (d, 1H), 7.09 (dd, 1H), 7.33 - 7.40 (m, 2H), 7.46 (d, 1H), 7.52 (d, 1H), 7.60 (d, 1H).

The procedures described in Method D i) - iii) were repeated using the appropriate benzyl halide or, using the alkyl indole-2-carboxylates as prepared by method F&G, with the appropriate benzyl halide. Thus were obtained the compounds described below.

Method D1.

Ethyl 5-acetoxy-N-[(2,3-dichloropyrid-5-yl)methyl]indole-2-carboxylate

90% yield. NMR: δ 1.27 (t, 3H), 2.26 (s, 3H), 4.28 (q, 2H), 5.85 (s, 2H), 7.12 (dd, 1H), 7.38 (s, 1H), 7.47 (d, 1H), 7.68 (d, 1H), 7.78 (d, 1H), 8.10 (d, 1H); m/z 409 (M+H⁺), 407.

Method D2.

Ethyl 5-acetoxy-N-(3-chloro-4-fluorobenzyl)indole-2-carboxylate

57% yield. NMR (CDCl₃): δ 1.37 (t, 3H), 2.33 (s, 3H), 4.35 (q, 2H), 5.74 (s, 2H), 6.90 (m, 1H), 7.00 (d, 1H), 7.05 (dd, 1H), 7.13 (dd, 1H), 7.26 (d, 1H), 7.36 (s, 1H), 7.22 (d, 1H).

Ethyl 5-acetoxy-N-(4-chloro-3-fluorobenzyl)indole-2-carboxylate

5 73% yield. m/z 390 (MH⁺).

Ethyl 5-acetoxy-N-(3-chlorobenzyl)indole-2-carboxylate

93% yield. m/z 372 (MH⁺).

Ethyl 5-acetoxy-N-(3-trifluoromethylbenzyl)indole-2-carboxylate

91% yield. m/z 406 (MH⁺).

10 Ethyl 5-acetoxy-N-(4-chlorobenzyl)indole-2-carboxylate

70% yield. m/z 372 (MH⁺).

Ethyl 5-acetoxy-3-bromo-N-(3, 4-dichlorobenzyl)indole-2-carboxylate

86% yield. m/z 486 (MH⁺).

Ethyl 5-acetoxy-4-bromo-N-(3, 4-dichlorobenzyl)indole-2-carboxylate

15 62% yield. NMR δ 1.40 (t, 3H), 2.39 (s, 3H), 4.38 (q, 2H), 5.77 (s, 2H), 6.82 (dd, 1H), 7.08 (d, 1H), 7.18 (s, 1H), 7.22 (d, 1H), 7.32 (d, 1H), 7.42 (s, 1H); m/z 486 (MH⁺).

Ethyl 5-acetoxy-N-(3, 4-dichlorobenzyl)-3-methylindole-2-carboxylate

79% yield. NMR δ 1.40 (t, 3H), 2.36 (s, 3H), 2.40 (s, 3H), 4.35 (q, 2H), 5.76 (s, 2H), 6.83 (dd, 1H), 7.00 (d, 1H), 7.10 (d, 1H), 7.19 (s, 1H), 7.30 (d, 1H), 7.40 (s, 1H); m/z

20 421 (M+H⁺).

Ethyl 5-acetoxy-N-(3, 4-dichlorobenzyl)-3-chloroindole-2-carboxylate

83% yield. NMR δ 1.25 (t, 3H), 2.25 (s, 3H), 4.3 (q, 2H), 5.8 (s, 2H), 6.9 (d, 1H), 7.2 (m, 1H), 7.4 (m, 2H), 7.5 (d, 1H), 7.7 (d, 1H); m/z 441.8 (M+H⁺)

25 **Method E**

Ethyl N-(3,4-dichlorobenzyl)-5-hydroxyindole-2-carboxylate

Sodium ethoxide (1.86 g) was added to a stirred solution of ethyl 5-acetoxy-N-(3,4-dichlorobenzyl)indole-2-carboxylate (5.55 g) in ethanol (50 ml) under an atmosphere of argon. The reaction was stirred at room temperature for 2 hours, then concentrated *in vacuo* and the residue acidified with aqueous hydrochloric acid (2 M) and extracted with dichloromethane. Combined organic extracts were

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washed with water, saturated aqueous sodium chloride solution and dried. The solvent was removed *in vacuo* and the residue was triturated with hexane / diethyl ether to yield the product as a white solid (3.17 g, 92%). NMR: δ 1.26 (t, 3H), 4.25 (q, 2H), 5.75 (s, 2H), 6.81 - 6.91 (m, 2H), 6.98 (d, 1H), 7.19 (s, 1H), 7.29 (d, 1H), 7.38 (d, 1H), 7.50 (d, 1H), 9.06 (s, 1H); m/z 364 (M+H⁺).

Method F

Ethyl 5-acetoxy-3-bromoindole-2-carboxylate

N-Bromosuccinimide (0.14 g) was added to a stirred solution of ethyl 5-acetoxyindole-2-carboxylate (0.2 g) in DMF (3.0 ml). The reaction was stirred for 4 hours, then poured into water. The resulting precipitate was filtered and dried *in vacuo* to give the title compound as a white powder (0.23 g, 87%). NMR δ 1.38 (t, 3H), 2.23 (s, 3H), 4.38 (q, 2H), 7.10 (dd, 1H), 7.23 (d, 1H), 7.50 (d, 1H), 12.28 (bs, 1H); m/z 326 (M⁺).

Method F1

Ethyl 5-acetoxy-3-chloroindole-2-carboxylate

A solution of ethyl 5-acetoxyindole-2-carboxylate (500mg) in dichloromethane (10ml) was stirred at room temperature in the presence of *N*-chlorosuccinimide (297mg) and potassium carbonate (279mg) overnight. The resulting precipitate was collected by filtration, washed with cold dichloromethane followed by water and dried under vacuum overnight to give the desired product as a white powder (425mg, 75%). NMR: δ 1.35 (t, 3H), 2.25 (s, 3H), 4.4 (q, 2H), 7.1 (d, 1H), 7.3 (s, 1H), 7.5 (d, 1H), 12.2 (s, 1H); m/z 281.9 (M+H⁺).

Method G

Ethyl 5-acetoxy-3-methylindole-2-carboxylate

(i) Ethyl 5-methoxy-3-methylindole-2-carboxylate

Concentrated sulphuric acid (1 ml) was added to a solution of 4-methoxyphenyl hydrazine hydrochloride (11.2 g) and 2-oxobutyric acid (8.72 g) in ethanol (250 ml), and the solution heated at reflux for 16 hours. The reaction was cooled, concentrated *in vacuo*, and the residue triturated with ethanol to give the title

compound as a white solid (8.8 g, 59%). NMR δ 1.36 (t, 3H), 3.76 (s, 3H), 4.30 (q, 2H), 6.88 (dd, 1H), 7.03 (d, 1H), 7.28 (d, 1H), 11.28 (bs, 1H); m/z 232 (M-H⁺).

(ii) Ethyl 5-acetoxy-3-methylindole-2-carboxylate

Boron tribromide (25 g) was added dropwise to a stirred solution of ethyl 5-methoxy-3-methylindole-2-carboxylate (2.0 g) in dry dichloromethane (250 ml) at -78°C under an atmosphere of argon. The reaction was allowed to warm to room temperature and stirred for a further 2 hours. The reaction was poured into ice / saturated aqueous sodium hydrogen carbonate solution with stirring and extracted with ethyl acetate. Combined organic extracts were washed with saturated aqueous sodium hydrogen carbonate solution, water, aqueous saturated sodium chloride solution and dried (MgSO₄). The solution was concentrated *in vacuo* and the residue dissolved in ethyl acetate. DMAP (20 mg) and acetic anhydride (0.5 ml) were added and the solution heated at reflux for 5 minutes. The reaction was cooled, concentrated *in vacuo* and the residue triturated with ether to give the title compound as a white powder (0.4 g, 18%). NMR δ 1.37 (t, 3H), 2.25 (s, 3H), 2.50 (s, 3H), 4.34 (q, 2H), 7.00 (dd, 1H), 7.37 (d, 1H), 7.40 (d, 1H), 11.52 (bs, 1H); m/z 260 (M-H⁺).

In a similar manner but starting with ethyl 4-bromo-5-methoxyindole-2-carboxylate was prepared:-

Ethyl 5-acetoxy-4-bromoindole-2-carboxylate NMR δ 1.42(t, 3H), 2.39(s, 3H), 4.42(q, 2H), 7.02(d, 1H), 7.23(s, 1H), 7.35(d, 1H), 9.22(bs, 1H): m/z 324,326 (M-H⁺).

Method H

Methyl-N-(3,4-dichlorobenzyl)-4-fluoro-5-hydroxyindole-2-carboxylate

(i) 2-Fluoro-3-benzoyloxy benzaldehyde

2-Fluoro-3-hydroxybenzaldehyde (16.49g) was dissolved in dimethylformamide (200ml) and stirred under an argon atmosphere. Sodium hydride was added (60% in mineral oil, 5.18g) and the mixture was stirred for 30 minutes. Benzyl bromide was added (16.8ml) and the mixture was stirred overnight. Reaction mixture was concentrated *in vacuo* and the resulting residue was partitioned between

diethyl ether (200ml) and water (200ml). Combined organic extracts were washed with water (400ml), dried (MgSO_4) and concentrated *in vacuo*. The residue was purified by flash column chromatography, using a gradient of 0-10% ethyl acetate/iso-hexane as eluent to give the product as a yellow solid (18.41g) ^1H NMR (DMSO-d_6) δ 5.20(s, 2H), 7.2-7.6(m, 8H), 10.21(s, 1H)

(ii) Methyl-2-azido-3-(2-fluoro-3-benzyloxyphenyl)propenoate

A mixture of methylazidoacetate (36.64g) and 2-Fluoro-3-benzyloxy benzaldehyde (18.32g) in methanol (250ml) was added dropwise, with stirring, over 1 hour to a mixture of sodium methoxide (17.20g) in methanol (100ml) at -25°C under a stream of argon. Mixture was left to stir for 20 minutes, allowed to warm to 5°C and stirred overnight.

Resulting precipitate was filtered, then washed sequentially with cold methanol, dilute solution of acetic acid in water and water. Resulting solid was dried under vacuum to give the product as a pale brown solid (16.70g) which was used without purification.

(iii) Methyl-4-fluoro-5-benzyloxvindole-2-carboxylate

A solution of methyl-2-azido-3-(2-fluoro-3-benzyloxyphenyl)propenoate (16.7g) in xylene (600ml) was added dropwise with stirring to refluxing xylene (2.4L) over 1 hour and then stirred for a further 20 minutes. The reaction mixture was concentrated *in vacuo* and purified by flash column chromatography, using a gradient of 0-100% ethyl acetate/iso-hexane as eluent to give the product as a yellow solid (12.93g) ^1H NMR (DMSO-d_6) δ 3.85(s, 3H), 5.15(s, 2H), 7.05-7.45(m, 8H), 12.06(s, 1H); $M/z(+)$ 300.4 (MH^+)

(iv) Methyl-N-(3,4-dichlorobenzyl)-4-fluoro-5-benzyloxvindole-2-carboxylate

Sodium hydride (60% in mineral oil, 589mg) was added to a solution of methyl-4-fluoro-5-benzyloxyindole-2-carboxylate (4g) in dimethylformamide (100ml) and the mixture was stirred under an argon atmosphere for 30 minutes. 3,4-

dichlorobenzyl chloride (2.22ml) was added and the mixture stirred overnight. The reaction mixture was concentrated *in vacuo* and the residue partitioned between diethyl ether (100ml) and water (100ml). Organic extracts were washed with water (100ml), dried (MgSO₄), concentrated *in vacuo* and purified by flash column chromatography, using iso-hexane followed by 5% ethyl acetate/iso-hexane as eluent, to give the product as a yellow crystalline solid (4.61g) ¹H NMR (DMSO-d₆) δ 3.80(s, 3H), 5.15(s, 2H), 5.80(s, 2H), 6.85(m, 1H), 7.25-7.52(m, 10H); *M/z*(+) 458.2 (*MH*⁺)

10 (v) Methyl-N-(3,4-dichlorobenzyl)-4-fluoro-5-hydroxyindole-2-carboxylate

A mixture of methyl-N-(3,4-dichlorobenzyl)-4-fluoro-5-benzyloxyindole-2-carboxylate (8.22g) and 5% Pd/C (200mg) in ethyl acetate (250ml) was stirred under a hydrogen atmosphere overnight, filtered through celite, concentrated *in vacuo* and purified by flash column chromatography using a gradient of 0-50% ethyl acetate/iso-hexane as eluent to give the product as a brown solid (6.18g) ¹H NMR (DMSO-d₆) δ 3.80(s, 3H), 5.75(s, 2H), 6.85(m, 1H), 7.00(t, 1H), 7.22(m, 2H), 7.30(m, 1H), 7.50(m, 1H), 9.33(s, 1H); *M/z*(-) 366.2 (*MH*⁻)

Method I

20 Ethyl-N-(3,4-dichlorobenzyl) 3-methoxy-5-hydroxyindole-2-carboxylate

(i) Ethyl 5-benzyloxy diazoindole-2-carboxylate

Sodium nitrite (6g) was added portionwise to a solution of ethyl 5-benzyloxyindole-2-carboxylate in ethyl acetate (40ml) and acetic acid (20ml). The mixture was stirred for 18 hours and then partitioned between ethyl acetate and water.

25 The organic extracts were washed with water, saturated aqueous sodium hydrogen carbonate and dried. The solvent was removed *in vacuo* and the resulting gum was titrated with diethyl ether to give the product as an orange powder (1.8g) NMR: δ 1.45 (t, 3H), 4.5 (q, 2H), 5.1 (s, 2H), 7.05 (m, 2H), 7.3 (m, 5H), 7.9 (d, 1H); *m/z* 322 (*M+H*⁺)

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(ii) Ethyl 3-methoxy-5-benzyloxy indole-2-carboxylate

Rhodium octanoate (300mg) was added to a stirred solution of ethyl 5-benzyloxy diazoindole-2-carboxylate (2.0g) in toluene (100ml) and methanol (10ml).

The mixture was refluxed under an inert atmosphere for 2.5 hours. The solution was concentrated *in vacuo* and the residue purified by column chromatography using 30 - 50% diethyl ether / iso-hexane to give an orange solid (1.41g). NMR: δ 1.4 (t, 3H), 4.05 (s, 3H), 4.4 (q, 2H), 5.1 (s, 2H), 7.1 (dd, 1H), 7.2-7.5 (m, 8H); m/z 326 (MH⁺)

5 (iii) Ethyl-N-(3,4-dichlorobenzyl) 3-methoxy-5-benzyloxy indole-2-carboxylate

3,4 Dichlorobenzyl chloride (0.72ml) was added to a stirred solution of ethyl 3-methoxy-5-benzyloxyindole-2-carboxylate (1.40g), potassium carbonate (0.90g) and potassium iodide (0.1g) in DMF (50ml) under inert atmosphere. The reaction mixture was heated to 50°C for 6 hours, then partitioned between ethyl acetate and water. Combined organic extracts were washed with water, then 3 times with saturated aqueous sodium chloride solution and dried. The solvent was removed *in vacuo* and the residue purified by column chromatography using 10 - 30% ethyl acetate / iso-hexane to give a yellow oil (0.9g). NMR: δ 1.4 (t, 3H), 4.0 (s, 3H), 4.4 (q, 10 2H), 5.1 (s, 2H), 5.6 (s, 2H), 6.9 (dd, 1H), 7.0-7.5 (m, 9H); m/z 484 (MH⁺)

15 (iv) Ethyl-N-(3,4-dichlorobenzyl) 3-methoxy-5-hydroxyindole-2-carboxylate

5% Pd/C (100mg) was added to a stirred solution of ethyl-N-(3,4-dichlorobenzyl) 3-methoxy-5-benzyloxyindole-2-carboxylate (0.9g) in ethyl acetate 20 (50ml) and the mixture was hydrogenated for 12 hours. Catalyst filtered off and filtrate evaporated to give a brown oil (0.61g) which was used without further purification. NMR: δ 1.4 (t, 3H), 4.0 (s, 3H), 4.4 (q, 2H), 5.6 (s, 2H), 6.8 (dd, 1H), 6.9 (dd, 1H), 7.1 (m, 3H), 7.3 (d, 1H); m/z 394 (MH⁺)

25 **Method J**

Ethyl N-(3,4-dichlorobenzyl)-4-chloro-5-methoxyindole-2-carboxylate

(i) Ethyl-2-azido-3-(2-chloro-3-methoxyphenyl)propenoate

30 A solution of ethyl azidoacetate (9.9 g) and 2-chloro-3-methoxybenzaldehyde (3 g) in ethanol (20 ml) was added dropwise to a solution of sodium ethoxide (4.7 g) in ethanol (10 ml) at 0°C. The reaction was allowed to warm to ambient temperature over 18 hours then partitioned between 2N HCl (50 ml) and dichloromethane

(250ml). The organic phase was dried (MgSO₄), concentrated under vacuo. and the residue purified by column chromatography using isohexane - 12% ethylacetate/isohexane as eluent to give the product as a pale yellow crystalline solid (2.2 g, 44%),

5 This was used without further purification

(ii) Ethyl-4-chloro-5-methoxyindole-2-carboxylate

A solution of ethyl-2-azido-3-(2-chloro-3-methoxyphenyl)propenoate (2.22 g) in xylene (100 ml) was heated at reflux for 30 minutes, concentrated *in vacuo* and the
10 residue purified by column chromatography using isohexane-50% ethyl acetate as eluent to give the product as a pale yellow solid (1.34 g, 67%). NMR δ (CDCl₃) 1.31 (t, 3H), 3.84 (s, 3H), 4.32 (q, 2H), 7.0 (d, 1H), 7.22 (d, 1H), 7.39 (d, 1H), 12.2 (bs, 1H);

(iii) Ethyl-N-(3,4-dichlorobenzyl)-4-chloro-5-methoxyindole-2-carboxylate

15 Sodium hydride (60mg) was added to a solution of ethyl-4-chloro-5-methoxyindole-2-carboxylate (250mg), 3,4-dichlorobenzylchloride (0.21ml) and tetrabutylammoniumiodide (3mg) in DMF at ambient temperature under an inert atmosphere. The reaction was stirred at ambient temperature for 18 hours then partitioned between ethylacetate (30ml) and water(30ml). The organic phase was dried
20 (MgSO₄), concentrated under vacuo and the residue purified by column chromatography using isohexane-15% ethylacetate as eluent to yield the product as a white solid (196mg, 48%) NMR: δ (CDCl₃) 1.39 (t, 3H), 3.93 (s, 3H), 4.32 (q, 2H), 5.73(s, 2H), 6.84 (dd, 1H), 7.06-7.16 (m, 3H), 7.31 (d, 1H), 7.42 (s, 1H)

25 (iv) Ethyl N-(3,4-dichlorobenzyl)-4-chloro-5-hydroxyindole-2-carboxylate

Trimethylsilyliodide (0.6ml) was added to a solution of ethyl-N-(3,4-dichlorobenzyl)-4-chloro-5-methoxyindole-2-carboxylate (190mg) in chloroform (20ml). The mixture was heated to 50 C for 18 hours then poured into methanol (50ml) and concentrated under vacuo. The residue was purified by column chromatography using isohexane-
30 20% ethyl acetate/isohexane as eluent to yield the product as a yellow solid (100mg, 71%) NMR: δ (CDCl₃) 1.39 (t, 3H), 4.35 (q, 2H), 5.72 (s, 2H), 6.84 (dd, 1H), 7.05-7.13 (m, 3H) 7.3 (d, 1H) 7.35 (s, 1H); m/z 396.2/398.2 (M-H⁺)

Example 15N-(3,4-Dichlorobenzyl)-2-trifluoromethylsulfonylamido-5-hydroxyindole

Sodium methoxide (21mg) was added to a stirred solution of N-(3,4-dichlorobenzyl)-2-trifluoromethylsulfonylamido-5-acetoxyindole (90mg) in methanol (10ml). Reaction stirred at ambient temperature for 1.5 hours then concentrated and acidified by the addition of aqueous hydrochloric acid (2M, 5ml), extracted with dichloromethane and concentrated *in vacuo* to give brown oil. (50mg).
NMR: δ 5.8 (s, 2H), 6.7 (dd, 1H), 6.9(m, 1H), 7.0(dd, 1H), 7.2 (dd, 1H), 7.3 (m, 1H), 7.5 (d, 1H); m/z 465, 467(M-H⁺).

The starting material for the above was prepared by :-

(i) N-(3,4-Dichlorobenzyl)-5-acetoxyindole-2-carboxylic acid

Dimethylaminopyridine (100mg) and acetic anhydride (1.12ml) were added to a solution of N-(3,4-Dichlorobenzyl)-5-hydroxyindole-2-carboxylic acid in ethyl acetate (50ml) and stirred at ambient temperature for 1 hour. Ethanol (10ml) was added and the reaction was stirred for 30min. Solvent partially evaporated and iso-hexane added to give a precipitate, which was filtered off and dried to give the product as a white solid (1.12g).
NMR: δ 2.25 (s, 3H), 5.85 (s, 2H), 6.9 (dd, 1H), 7.3 - 7.6 (m, 5H); m/z 376, 378 (M-H⁺)

(ii) N-(3,4-Dichlorobenzyl)-2-trifluoromethylsulfonylamido-5-acetoxyindole

To a stirred solution of N-(3,4-Dichlorobenzyl)-5-acetoxyindole-2-carboxylic acid in DMF (5ml), under an inert atmosphere, was added HATU (0.27g), DIPEA (0.12ml) and trifluoromethylsulfonamide (97mg). Reaction was stirred at ambient temperature for 18 hours. The mixture was poured into saturated sodium bicarbonate solution and the resulting precipitate was filtered off and dried to give product. (90mg). NMR: δ 2.25 (s, 3H), 5.9 (s, 2H), 6.9 (dd, 1H), 7.0(dd, 1H), 7.1(s, 1H), 7.35 (m, 1H); m/z 506, 508(M-H⁺)

Example 16N-(3,4-Dichlorobenzyl)-5-hydroxyindole-2-tetrazole

Ammonium chloride (54mg) and sodium azide (65mg) were added to a stirred solution of N-(3,4-Dichlorobenzyl)-5-acetoxyindole-2-nitrile in DMF (5ml). The reaction mixture was heated to 100°C for 10 hours. A further amount of ammonium chloride (35mg) and sodium azide (42mg) was added and the reaction heated to 100°C for 18 hours. The reaction mixture was acidified by the addition of aqueous hydrochloric acid (2M, 10ml) and extracted with ethyl acetate, dried, concentrated *in vacuo* and purified by column chromatography using 20% ethyl acetate in iso-hexane, increasing to 5% methanol in ethyl acetate to give the product as a brown oil (40mg) which solidified on standing.

NMR: δ 5.9 (s, 2H), 6.75 (dd, 1H), 6.9 (dd, 1H), 7.1(s,1H), 7.1-7.3 (m,2H), 7.5 (d, 1H), 9.0 (s,1H); m/z 360/362 (MH⁺)

The starting material was prepared by :-

Methane sulfonyl chloride (0.5ml) was added to a cooled (0°C) solution of N-(3,4-dichlorobenzyl)-5-acetoxyindole-2-carboxylic acid (1.12g) in pyridine (30ml) and stirred at 0°C for 1.5 hours. Gaseous ammonia was bubbled through the reaction mixture for 15 min, then excess ammonia removed *in vacuo*. Reaction mixture cooled to 0°C and methyl sulfonyl chloride (2.5ml) added to the stirred solution and allowed to reach ambient temperature over 18 hours. Methane sulfonyl chloride (2ml) added and reaction mixture left to stand for 60 hours. The solvent was removed *in vacuo*, re-dissolved in dichloromethane and washed 3 times with a 1:1 mixture of aqueous hydrochloric acid (1M) and saturated ammonium chloride solution. The organic extracts were dried, concentrated *in vacuo*, and the residue purified by column chromatography, using 10 - 25% ethyl acetate / iso-hexane to give the desired product. (300mg). NMR: δ 2.25 (s, 3H), 5.6 (s, 2H), 7.0 (dd, 1H), 7.45 - 7.65 (m, 4H), 7.7 (d, 1H).

Example 17N-(3,4-dichlorophenylsulphonyl)-5-hydroxyindole-2-carboxylic acid

A solution of anhydrous lithium iodide (870mg) and methyl N-(3,4-dichlorophenylsulphonyl)-5-hydroxyindole-2-carboxylate (260mg) in pyridine (15ml) was stirred at reflux for 4 hours. The reaction was cooled and concentrated *in vacuo*. The residue was dissolved in water (20ml) and acidified with acetic acid. The product was extracted with ethyl acetate and the combined extracts were dried, concentrated *in vacuo* and the residue purified by column chromatography using dichloromethane-50% ethyl acetate containing 1% acetic acid as eluent to give the desired product as a glass (72mg, 29%). NMR: δ 6.9 (m, 2H), 7.25 (s, 1H), 7.85 (d, 1H), 7.9 (m, 2H), 8.15 (s, 1H), 9.5 (s, 1H); m/z 385.8 (M-H⁺).

The starting material was prepared by

(i) Methyl N-(3,4-dichlorophenylsulphonyl)-5-benzyloxyindole-2-carboxylate

Sodium Hydride (60% dispersion, 444mg) was added to a stirred solution of methyl 5-benzyloxyindole-2-carboxylate (2.08g) in DMF (50ml) at room temperature. After 1 hour, 3,4-dichlorobenzenesulphonyl chloride (2.72g) was added. Stirring was continued for 2 hours after which the reaction mixture was partitioned between water and ethyl acetate. Combined organic extracts were dried and concentrated *in vacuo* and the residue purified by column chromatography using isohexane-20% ethyl acetate as eluent to give the desired product as a white solid (2.02g, 56%). NMR: δ 3.85 (s, 3H), 5.1 (s, 2H), 7.2 (m, 1H), 7.4 (m, 7H), 7.9 (s, 2H), 8.0 (d, 1H), 8.2 (s, 1H); m/z 489.8 (MH⁺).

(ii) Methyl N-(3,4-dichlorophenylsulphonyl)-5-hydroxyindole-2-carboxylate

A suspension of 5% palladium on carbon in ethyl acetate (450ml) and methyl N-(3,4-dichlorophenylsulphonyl)-5-benzyloxyindole-2-carboxylate (2.01g) was stirred at 60°C under hydrogen at atmospheric pressure for 48 hours. The catalyst was removed by filtration and the filtrate concentrated *in vacuo*. The residue was purified by column chromatography using 20% ethyl acetate / isohexane as eluent to give the desired product as a gum (270mg, 16%). NMR: δ 3.85 (s, 3H), 7.0 (m, 2H), 7.35 (s, 1H), 7.9 (m, 3H), 8.1 (s, 1H), 9.6 (s, 1H); m/z 401.9 (MH⁺).

Example 18*N*-(3,4-Dichlorobenzyl)-5-acetoxyindole-2-carboxylic acid

To a solution of *N*-(3,4-Dichlorobenzyl)-5-hydroxyindole-2-carboxylic acid (10g) in warm ethyl acetate (250ml) was added 4-dimethylaminopyridine (100mg) and acetic anhydride (5.0ml) and the resulting mixture was stirred for 2 hours. The organics were washed with 1N HCl and dried. Hexane was added to cause crystallisation of the product. The solid was filtered and washed with hexane to give the desired product. (5g, 44%). ¹H NMR (DMSO-d₆) δ 2.25 (s, 3H), 5.85 (s, 2H), 6.9 (dd, 1H), 7.05 (dd, 1H), 7.3-7.6 (m, 5H); m/z 378, 380 (MH⁺).

Example 19**Pharmaceutical Compositions**

This Example illustrates, but is not intended to limit, representative pharmaceutical dosage forms of the invention as defined herein (the active ingredient being termed "Compound X"), for therapeutic or prophylactic use in humans:

Example A

(a)

<u>Tablet I</u>	<u>mg/tablet</u>
Compound X.	100
Lactose Ph.Eur	182.75
Croscarmellose sodium	12.0
Maize starch paste (5% w/v paste)	2.25
Magnesium stearate	3.0

(b)

<u>Tablet II</u>	<u>mg/tablet</u>
Compound X	50
Lactose Ph.Eur	223.75
Croscarmellose sodium	6.0
Maize starch	15.0
Polyvinylpyrrolidone (5% w/v paste)	2.25
Magnesium stearate	3.0

(c)

<u>Tablet III</u>	<u>mg/tablet</u>
Compound X	1.0
Lactose Ph.Eur	93.25
Croscarmellose sodium	4.0
Maize starch paste (5% w/v paste)	0.75
Magnesium stearate	1.0

(d)

<u>Capsule</u>	<u>mg/capsule</u>
Compound X	10
Lactose Ph.Eur	488.5
Magnesium	1.5

5

(e)

<u>Injection I</u>	<u>(50 mg/ml)</u>
Compound X	5.0% w/v
1M Sodium hydroxide solution	15.0% v/v
0.1M Hydrochloric acid	to adjust pH to 7.6
Polyethylene glycol 400	4.5% w/v
Water for injection	to 100%

(f)

<u>Injection II</u>	<u>(10 mg/ml)</u>
Compound X	1.0% w/v
Sodium phosphate BP	3.6% w/v
0.1M Sodium hydroxide solution	15.0% v/v
Water for injection	to 100%

(g)

<u>Injection III</u>	(1mg/ml, buffered to pH6)
Compound X	0.1% w/v
Sodium phosphate BP	2.26% w/v
Citric acid	0.38% w/v
Polyethylene glycol 400	3.5% w/v
Water for injection	to 100%

(h)

<u>Aerosol I</u>	<u>mg/ml</u>
Compound X	10.0
Sorbitan trioleate	13.5
Trichlorofluoromethane	910.0
Dichlorodifluoromethane	490.0

5 (i)

<u>Aerosol II</u>	<u>mg/ml</u>
Compound X	0.2
Sorbitan trioleate	0.27
Trichlorofluoromethane	70.0
Dichlorodifluoromethane	280.0
Dichlorotetrafluoroethane	1094.0

(j)

<u>Aerosol III</u>	<u>mg/ml</u>
Compound X	2.5
Sorbitan trioleate	3.38
Trichlorofluoromethane	67.5
Dichlorodifluoromethane	1086.0
Dichlorotetrafluoroethane	191.6

(k)

<u>Aerosol IV</u>	<u>mg/ml</u>
Compound X	2.5
Soya lecithin	2.7
Trichlorofluoromethane	67.5
Dichlorodifluoromethane	1086.0
Dichlorotetrafluoroethane	191.6

(l)

<u>Ointment</u>	<u>ml</u>
Compound X	40 mg
Ethanol	300 μ l
Water	300 μ l
1-Dodecylazacycloheptan-2-one	50 μ l
Propylene glycol	to 1 ml

5

Note:

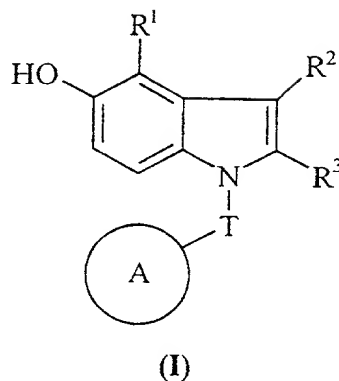
Compound X in the above formulations may comprise a compound as illustrated in Examples 1 to 3 herein.

The above formulations may be obtained by conventional procedures well known in the pharmaceutical art. The tablets (a)-(c) may be enteric coated by conventional means, for example to provide a coating of cellulose acetate phthalate. The aerosol formulations (h)-(k) may be used in conjunction with standard, metered dose aerosol dispensers, and the suspending agents sorbitan trioleate and soya lecithin may be replaced by an alternative suspending agent such as sorbitan monooleate, sorbitan sesquioleate, polysorbate 80, polyglycerol oleate or oleic acid.

15

Claims

1. A compound of the formula (I):



wherein:

R¹ is hydrogen, halo or methoxy;

R² is hydrogen, halo, methyl, ethyl or methoxy;

R³ is carboxy, tetrazolyl or -CONHSO₂R⁴ where R⁴ is methyl, ethyl, phenyl,
2,5-dimethylisoxazolyl or trifluoromethyl;

T is -CH₂- or -SO₂-; and

ring A is 3-chlorophenyl, 4-chlorophenyl, 3-trifluoromethylphenyl,

3,4-dichlorophenyl, 3,4-difluorophenyl, 3-fluoro-4-chlorophenyl,

3-chloro-4-fluorophenyl or 2,3-dichloropyrid-5-yl;

or a pharmaceutically acceptable salt or prodrug thereof.

2. A compound according to claim 1 wherein ring A is 3-chlorophenyl,
4-chlorophenyl, 3-trifluoromethylphenyl, 3,4-dichlorophenyl, 3,4-difluorophenyl,
3-fluoro-4-chlorophenyl or 3-chloro-4-fluorophenyl.

3. A compound according to claim 2 wherein ring A is 3,4-dichlorophenyl,
3-fluoro-4-chlorophenyl or 3-chloro-4-fluorophenyl.

4. A compound according to claim 1 wherein ring A is 3,4-dichlorophenyl,
2,3-dichloropyrid-5-yl or 3-chloro-4-fluorophenyl.

5. A compound according to any one of the preceding claims wherein T is $-\text{CH}_2-$.

6. A compound according to any one of the preceding claims where R^3 is carboxy.

5

7. A compound according to claim 1 wherein in the compound of formula (I),

R^1 is hydrogen;

R^2 is hydrogen;

R^3 is carboxy;

10

T is $-\text{CH}_2-$; and

ring A is 3,4-dichlorophenyl or 3-chloro-4-fluorophenyl;

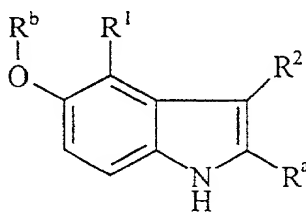
or a pharmaceutically acceptable salt or prodrug thereof.

8. A process for preparing a compound according to claim 1 which process

15

comprises:

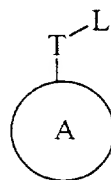
a) reacting compounds of formula (II):



(II)

where R^a is a group R^3 as defined in claim 1 or protected form of a group R^3 , R^b is

20 hydrogen or a hydroxy protecting group, and R^1 and R^2 are as defined in claim 1, with a compound of formula (III):



(III)

where T and ring A are as defined in claim 1, and L is a displaceable group;

25 and thereafter if necessary:

- i) converting a compound of the formula (I) into another compound of the formula (I);
- ii) removing any protecting groups; or
- iii) forming a pharmaceutically acceptable salt or prodrug thereof.

5 9. A pharmaceutical composition comprising a compound according to any one of claims 1 to 7 in combination with a pharmaceutically acceptable carrier.

10 10. A compound according to any one of claims 1 to 7 for use in the preparation of a medicament for use in the treatment of disease mediated by monocyte chemoattractant protein-1 or RANTES (Regulated upon Activation, Normal T-cell Expressed and Secreted), such as inflammatory disease.

FOR UTILITY/DESIGN
CIP/PCT NATIONAL/PLANT
ORIGINAL/SUBSTITUTE/SUPPLEMENTAL
DECLARATIONS

RULE 63 (37 C.F.R. 1.63)
DECLARATION AND POWER OF ATTORNEY
FOR PATENT APPLICATION
IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

PW
FORM

Z70472/UST

As a below named inventor, I hereby declare that my residence, post office address and citizenship are as stated below next to my name, and I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the INVENTION ENTITLED: INDOLE DERIVATIVES AND THEIR USE AS MCP-1 INHIBITORS

the specification of which (CHECK applicable BOX(ES))
X A ☐ is attached hereto.
BOX(ES) → B. ☐ was filed on _____ as U.S. Application No. _____
→ C. ☐ was filed as PCT International Application No. PCT/GB00/00265 on 31 January 2000
and (if applicable to U.S. or PCT application) was amended on _____ I hereby state that I have reviewed and understand the contents of the

above identified specification including the claims, as amended by any amendment referred to above. I acknowledge the duty to disclose all information known to me to be material to patentability as defined in 37 C.F.R. 1.56. Except as noted below, I hereby claim foreign priority benefits under 35 U.S.C. 119(a)-(d) or 365(b) of any foreign application(s) for patent or inventor's certificate or 365(a) of any PCT International Application which designated at least one other country than the United States, listed below and have also identified below any foreign application for patent or inventor's certificate, or PCT International Application, filed by me or my assignee disclosing the subject matter claimed in this application and having a filing date (1) before that of the application on which priority is claimed or (2) if no priority claimed, before the filing date of this application

<u>PRIOR FOREIGN APPLICATION(S)</u> <u>Number</u>	<u>Country</u>	<u>Date first laid-</u> <u>open or Published</u>	<u>Date Patented</u> <u>or Granted</u>	<u>Priority NOT Claimed</u>
9902461.4	United Kingdom	05.02.1999		

Except as noted below, I hereby claim domestic priority benefit under 35 U.S.C. 119(e) or 120 and/or 365(c) of the indicated United States applications listed below and PCT international applications listed above or below and, if this is a continuation-in-part (CIP) application, insofar as the subject matter disclosed and claimed in this application is in addition to that disclosed in such prior applications, I acknowledge the duty to disclose all information known to me to be material to patentability as defined in 37 C.F.R. 1.56 which became available between the filing date of each such prior application and the national or PCT international filing date of this application:

<u>PRIOR U.S. PROVISIONAL, NON PROVISIONAL AND/OR PCT APPLICATION(S)</u> <u>Application No. (series code/serial no.)</u>	<u>Date/MONTH/Year Filed</u>	<u>Status</u> <u>Pending, abandoned, patented</u>	<u>Priority NOT Claimed</u>
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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true, and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon

And I hereby appoint Pillsbury Winthrop LLP, Intellectual Property Group, telephone number (202)861-3000 (to whom all communications are to be directed), and persons of that firm who are associated with USPTO Customer No.909 (see below label) individually and collectively my attorneys to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith and with the resulting patent, and I hereby authorize them to delete from that Customer No. names of persons no longer with their firm, to add new persons of their firm to that Customer No., and to act and rely on instructions from and communicate directly with the person/assignee/attorney/firm/organization who/which first sends/sent this case to them and by whom/which I hereby declare that I have consented after full disclosure to be represented unless/until I instruct the above firm and/or an attorney of that firm in writing to the contrary

00909

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☐ OR ADDITIONAL INVENTORS see attached page.

☐ See additional foreign priorities on attached page (incorporated herein by reference).

Atty. Dkt. No. P (M#)

DECLARATION AND POWER OF ATTORNEY
(continued)

Additional Inventors

Page 2

(3) INVENTOR'S SIGNATURE

Date: